

An Investigation of the Ability of the Glutaraldehyde Test to Distinguish between Acute and Chronic Inflammatory Disease in Horses

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Brink P, Wright JC, Schumacher J: An investigation of the ability of the glutaraldehyde test to distinguish between acute and chronic inflammatory disease in horses. Acta vet. scand. 2005, 46, 69-78. – The glutaraldehyde test (GT), a rapid and inexpensive test, has been utilized empirically for many years in bovine practice for diagnosing inflammatory diseases. GT is used primarily to demonstrate increased serum concentrations of fibrinogen and globulin. Glutaraldehyde binds with free amino groups in fibrinogen and immunoglobulin to create a clot in a first degree chemical reaction. The clotting time of the GT estimates the content of proteins produced in response to inflammation. The applicability of GT for diagnosing inflammation in the horse has never been investigated. The objective of this study was to determine the ability of GT to distinguish between acute and chronic inflammatory disease in horses. Thirty-seven horses with suspected inflammatory diseases were evaluated using the GT, history, complete clinical examination and routine blood analysis. GT-times, laboratory results and clinical outcome were compared statistically. Horses that were determined to be acutely affected (based on history, clinical examination and routine blood analysis) tended to have a negative GT (75%). Results of the GT did not correlate with blood fibrinogen concentration. Positive GT also predicted a fatal outcome in 69% of the clinical cases. The results of this trial indicate that GT can be a useful screening test to distinguish between acute and chronic inflammatory disease in horses.

Glutaraldehyde test, inflammation, horse diseases, equine, diagnostic techniques, prognosis, immunoglobulin, globulin, blood clot, infectious diseases, hypergamma-globulinemia, serum biochemistries.

Introduction

The glutaraldehyde reagent in the glutaraldehyde test (GT) creates a clot with either fibrinogen or gammaglobulin in EDTA-stabilized blood by chemical reaction between the aldehyde groups in glutaraldehyde and free amino groups in fibrinogen and immunoglobulins (Sandholm 1974a, Sandholm 1974b, Martin et al. 1985). The process is believed to run as a first degree chemical reaction, where the reac-

tion time is directly proportional to the concentration of fibrinogen and immunoglobulins (Sandholm 1974a, Sandholm 1974b, Eriksen 1984).

The rapid and inexpensive GT has been used with success empirically in Europe for many years for diagnosing inflammatory diseases in cattle (Sandholm 1974a, Sandholm 1974b, Liberg et al. 1975a, Liberg et al. 1975b, Nielsen

1975, Martens 1977, Liberg 1978, Tennant et al. 1979, Liberg 1981, Liberg 1982, Eriksen 1984, Keulen et al. 1984, Doll et al. 1985, Larsson 1985, Mahlin et al. 1985, Chadli & Mahin 1986, Kovac 1988, Katholm & Jorgensen 1992, Kantor et al. 1993, Tyler et al. 1996, Sen et al. 2000, Ramprabhu et al. 2002), pigs (Liberg 1979, Hansen 1985, Kovac et al. 1993), goats (Satpathy et al. 1996, Vihan 1989), mink (Sandholm & Kangas 1973), dogs (Sandholm & Kivistö 1975, Wolff 1986), and zoo animals (O'Rourke & Satterfield 1981, Carstairs-Grant et al. 1988, Juyal & Uppal 1995). In these species, the test was used to indicate whether an inflammatory disease was acute or chronic (Doll et al. 1985, Chadli & Mahin 1986).

The GT, because of its simplicity, is very useful in bovine practice for rapidly diagnosing inflammation under circumstances where it is not practical or economically possible to have blood analyzed at a professional clinical laboratory (Sandholm 1974a, Sandholm 1974b, Liberg et al. 1975a, Liberg et al. 1975b, Nielsen 1975, Martens 1977, Liberg 1978, Tennant et al. 1979, Liberg 1981, Liberg 1982, Eriksen 1984, Keulen et al. 1984, Doll et al. 1985, Larsson 1985, Mahlin et al. 1985, Chadli & Mahin 1986, Kovac 1988, Katholm & Jorgensen 1992, Kantor et al. 1993, Tyler et al. 1996, Sen et al. 2000, Ramprabhu et al. 2002).

A negative GT can be used as a semiquantitative indicator of hypogammaglobulinemia caused by failure of passive transfer of colostrum in neonatal foals (Beetson et al. 1985, Clabough et al. 1989, Saikku et al. 1989, Clabough et al. 1991, Kumaran & Bhuvanakumar 1994, Kalinbacak & Or 1996, Bruijn et al. 2003), calves (Tennant et al. 1979, Keulen et al. 1984, Larsson 1985, Kovac 1988, Tyler et al. 1996, Sen et al. 2000), kids (Vihan 1989, Satpathy et al. 1996), and zoo ruminants (O'Rourke & Satterfield 1981, Carstairs-Grant et al. 1988, Juyal & Uppal 1995). The

GT also has been used to determine the content of IgG in mare colostrum (Jones & Brook 1995, Ezhilan & Bhuvanakumar 1998).

Clinical experience indicates that the GT may not be as reliable in horses as it is in cattle (Nielsen 1975). In horses, lack of reliability of the GT has been proposed to be caused by generally lower or delayed peaks of concentrations of fibrinogen and immunoglobulin or a different distribution of immunoglobulins (IgG, IgM, IgA) compared to cattle (Bendixen 1954, Nansen & Nielsen 1966, Sandholm 1974a, Nielsen 1975, Aasted et al. 1989).

The purpose of this clinical trial was to determine the ability of GT to distinguish between acute and chronic inflammatory disease in horses. During the trial we compared indicators of inflammation (the concentration of blood fibrinogen and serum globulin) to the GT.

Materials and methods

Thirty seven horses admitted for investigation of suspected inflammatory disease were evaluated using the GT (Glutarvac^a), a complete clinical examination, CBC and routine serum biochemistries that included total protein, albumin, globulin and fibrinogen. Blood for the GT and laboratory analysis was collected at the same time either upon arrival at the hospital or the following day.

Horses having a history of clinical signs of inflammatory disease of total duration six days or less were arbitrarily classified as acutely inflamed. Horses with a history of clinical signs greater than six days were arbitrarily classified as chronically inflamed. The clinical examination leading to the diagnosis and etiology was also used to reinforce the distinction between acute and chronic disease (Table 1).

The GT was performed by adding equal amounts of fresh blood and glutaraldehyde in a test tube, mixing by slowly turning the test tube and visually observing and noting the time re-

Table 1: Diagnosis and outcome.

Horse #	Diagnosis	Duration	Outcome
1	Purulent, bilateral guttural pouch empyema	Chronic	Fatal (spontaneous)
2	Dorsal rectal abscess	Chronic	Discharged
3	Traumatic, infected joint capsular laceration	Acute	Discharged
4	Dorsal rectal abscesses	Chronic	Discharged
5	Purulent nephritis, lung abscesses, ulcerous dermatitis, myocarditis, fatty liver	Chronic	Fatal (euthanasia)
6	Severe, idiopathic, systemic infection	Acute	Fatal (spontaneous)
7	Purulent (jugular) thrombophlebitis (abscess)	Chronic	Discharged
8	Transportation syndrome, bronchitis/pleuritis, systemic infection	Acute	Discharged
9	Fibrinopurulent pleuropneumonia	Acute	Fatal (euthanasia)
10	Systemic, malign lymphoma, borrelia infection	Chronic	Fatal (euthanasia)
11	Infected tendovaginitis	Acute	Discharged
12	Intraabdominal abscess, squamous cell carcinoma (ventricle)	Chronic	Fatal (euthanasia)
13	Septic, purulent arthritis	Chronic	Discharged
14	Fibrinopurulent pleuropneumonia	Acute	Fatal (euthanasia)
15	Septicemia, pneumonia, peritonitis	Acute	Fatal (euthanasia)
16	Severe, purulent, traumatic muscle laceration	Chronic	Discharged
17	Severe, iatrogenic, muscle abscesses	Chronic	Discharged
18	Purulent osteomyelitis	Chronic	Fatal (euthanasia)
19	Severe subcutaneous infection/abscess, funiculitis	Chronic	Discharged
20	Humerus fracture, subcutaneous infection/abscess	Chronic	Discharged
21	Bacterial diarrhea	Acute	Fatal (euthanasia)
22	Abscess, inguinal region	Chronic	Discharged
23	Scrotal abscesses, postoperative castration	Chronic	Discharged
24	Necrotizing myositis, multiple subcutaneous abscesses	Chronic	Discharged
25	Fibrinopurulent septic bicipital bursitis, muscular septic cellulitis	Chronic	Fatal (euthanasia)
26	Pericarditis, mitral insufficiency, systemic infection	Chronic	Fatal (euthanasia)
27	Septic peritonitis	Chronic	Discharged
28	Septic meningitis	Acute	Discharged
29	Septicemia, premature foal	Acute	Discharged
30	M. Masseter, throat latch, parotid, jugular abscesses/fistulae	Chronic	Discharged
31	Systemic infection, septic myositis	Chronic	Fatal (euthanasia)
32	Systemic infection, possible abdominal/kidney abscess, emaciation	Chronic	Fatal (euthanasia)
33	Severe, multiple, purulent, septic arthritis	Chronic	Fatal (euthanasia)
34	Metritis, purulent peritonitis, abdominal abscesses, adherences	Chronic	Fatal (euthanasia)
35	Purulent, pharyngeal inflammation, choke	Acute	Discharged
36	Thrombosis pulmonary vessels, Cushing disease, laminitis	Chronic	Fatal (euthanasia)
37	Systemic intoxication, parasitic aneurysm, intestinal volvulus, paralysis	Acute	Fatal (euthanasia)

Table 2: Categorization of GT-time.

Group #	GT-times	Empiric categorization
1	0 < GT-time < 3 min.	High increase in concentration of fibrinogen and/or immunoglobulin
2	3 < GT-time < 6 min.	Moderate increase in concentration of fibrinogen and/or immunoglobulin
3	6 < GT-time < 15 min.	Low increase in concentration of fibrinogen and/or immunoglobulin
4	GT >15 min.	No increase in concentration of fibrinogen and/or immunoglobulin

Table 3: Blood values

Horse #	GT-time	Albumin (min)	Globulin (g/l)	Alb/Glob (g/l)	Fibrinogen (ratio)	Total prot. (g/l)	WBC (10.9/l) (g/l)	Differential cell count leukocytes						RBC (10.12/l)	Hemoglobin (g/l)	PCV (%)
								Bands (%)	Segm (%)	Eosin (%)	Mono (%)	Lymph (%)	Baso (%)			
1	2,0	30	58	0,5	7,1	88	9,4	1	63	0	4	32	0	8,8	113	31
2	NR	30	35	0,9	7,5	65	9,3	0	42	0	5	53	0	9,7	118	33
3	NR	38	30	1,3	3,6	68	9,4	0	67	1	1	31	0	8,5	123	34
4	NR	27	35	0,8	4,7	62	7,0	3	24	2	3	66	2	8,2	110	30
5	NR	41	33	1,2	2,3	74	16,7	0	85	0	5	9	1	11,6	178	48
6	3,5	31	40	0,8	8,3	71	10,2	2	79	0	4	15	0	7,3	117	33
7	NR	36	37	1,0	7,7	73	16,0	5	78	0	1	16	0	11,7	186	50
8	NR	34	42	0,8	4,4	76	9,3	2	68	1	3	25	1	7,3	125	35
9	NR	32	37	0,9	9,9	69	9,2	11	64	0	4	21	0	8,6	144	39
10	1,0	17	63	0,3	3,0	80	15,8	0	80	1	4	14	1	1,1	27	8
11	NR	31	48	0,6	8,2	79	6,2	0	61	1	4	34	0	5,9	102	28
12	6,0	32	60	0,5	5,2	92	7,0	0	74	1	5	20	0	7,0	130	34
13	NR	34	29	1,2	5,9	63	11,1	0	68	0	3	29	0	8,0	105	31
14	5,0	18	47	0,4	7,1	65	10,3	3	59	0	8	30	0	9,2	153	45
15	NR	23	27	0,9	6,4	50	2,8	7	11	0	5	77	0	12,8	176	50
16	NR	33	25	1,3	2,9	58	7,4	1	68	6	1	23	1	7,8	136	38
17	NR	31	29	1,1	5,4	60	7,3	0	56	2	6	36	0	10,1	168	48
18	NR	27	25	1,1	4,9	52	35,4	0	94	0	3	3	0	6,5	137	39
19	3,0	29	58	0,5	6,8	87	19,2	1	72	2	1	24	0	5,8	85	26
20	NR	33	20	1,7	12,2	53	15,0	0	75	0	6	19	0	9,1	121	35
21	NR	20	18	1,1	7,0	38	36,6	0	90	0	1	9	0	10,3	136	37
22	3,0	21	69	0,3	5,0	90	30,7	0	85	1	1	13	0	6,3	92	24
23	NR	21	36	0,6	6,0	57	7,5	0	54	2	2	41	1	6,3	107	28
24	NR	21	19	1,1	5,0	40	13,7	1	87	0	2	10	0	5,5	72	19
25	NR	36	31	1,2	6,6	67	8,6	0	77	0	8	14	0	5,6	101	27
26	NR	35	22	1,6	6,0	57	10,8	0	86	2	2	10	0	8,3	141	41
27	5,0	31	46	0,7	9,0	77	13,5	0	72	0	3	25	0	7,4	119	32
28	NR	25	41	0,6	10,0	66	33,4	0	93	0	6	1	0	10,5	126	32
29	NR	32	16	2,0	5,0	48	0,8	0	16	0	0	84	0	7,7	117	31
30	NR	28	21	1,3	14,7	49	26,4	0	72	0	9	18	1	8,1	98	25
31	15,0	40	27	1,5	7,3	67	11,5	1	78	1	2	17	1	6,8	118	31
32	NR	36	19	1,9	4,6	55	15,0	2	42	2	7	47	0	9,3	116	32
33	15,0	29	44	0,7	4,8	73	8,0	1	45	0	9	45	0	8,0	128	35
34	14,0	21	23	0,9	5,8	44	9,4	0	90	2	2	6	0	6,2	114	31
35	2,0	31	42	0,7	11,0	73	12,0	0	75	1	2	21	1	5,1	83	20
36	3,0	32	41	0,8	5,0	73	16,7	0	93	0	2	5	0	4,3	80	20
37	NR	33	39	0,8	6,3	72	11,9	0	76	0	7	17	0	11,7	185	53

* NR = no reaction

Table 4: GT result versus mean blood values (+/- standard deviation).

	Albumin (g/l)	Globulin (g/l)	Alb/Glo (ratio)	Fibrinogen (g/l)
GT-positive	27,9 (+/- 6,6)	47,5 (+/- 13,7)	0,7 (+/- 0,3)	6,6 (+/- 2,1)
GT-negative	30,7 (+/- 5,7)	29,3 (+/- 8,7)	1,1 (+/- 0,4)	6,6 (+/- 2,9)
All horses	29,7 (+/- 6,1)	36,0 (+/- 13,6)	1,0 (+/- 0,4)	6,6 (+/- 2,6)

Table 5: Clinical parameters versus mean blood values.

	GT-positive (%)	Albumin (g/l)	Globulin (g/l)	Alb/Glo (ratio)	Fibrinogen (g/l)
Duration	Acute	23,1	26,7	43,0	0,6
	Chronic	76,9	28,2	48,9	0,7
Outcome	Fatal	69,2	27,8	44,8	0,7
	Discharged	30,8	28,0	53,8	0,6

Table 6: GT-time groups versus mean blood values within groups (+/- standard deviation).

Group #	GT-positive (No)	Mean Globulin (g/l)	Mean Alb/Glo (ratio)	Mean Fibrinogen (g/l)
1: (0<GT-time<3 min.)	6	55,2 (+/- 11,3)	0,5 (+/- 0,2)	6,3 (+/- 2,7)
2: (3<GT-time<6 min.)	4	48,3 (+/- 8,4)	0,6 (+/- 0,2)	7,4 (+/- 1,7)
3: (6<GT-time<15 min.)	3	31,3 (+/- 11,2)	1,0 (+/- 0,4)	6,0 (+/- 2,0)
4: (GT-time>15 min.)	24	29,3 (+/- 8,7)	1,1 (+/- 0,4)	6,6 (+/- 2,9)

quired for full clot formation. The test result was categorized respectively as high, moderate, low or no increase in concentration of fibrinogen and/or immunoglobulin based on GT-time (Table 2).

The results of the GT and fibrinogen, globulin and albumin/globulin ratio were compared using regression and correlation. The association of the GT results with fatality was analyzed using chi-square. All data from the blood analysis were also tested for correlation with GT using principal component analysis.

Results

In Table 1, diagnoses, estimated duration of the diseases and outcome of the clinical cases are summarized.

In Table 3, the GT-times and results of blood analysis of the horses are summarized.

Table 4 shows the mean concentration of se-

lected blood values for horses whose blood had positive reaction to the GT, compared to horses whose blood had a negative reaction to the GT. Table 5 shows the comparison of selected clinical parameters and mean blood values of horses with positive GT.

The GT-times were divided into groups as listed in Table 6. Table 7 shows the correlation of GT-time and Group number versus globulin concentration and albumin/globulin ratio, respectively, by linear regression. The regression equations are also shown in Graphs 1-4. Group number did not correlate with the mean fibrinogen concentration within groups.

Among the hospitalized horses, there was a higher fatality rate in the GT positive horses (69% = 9/13) when compared to the GT negative horses (38% = 9/24); however, this finding was not statistically significant ($p=0.06$, Chi square test).

Table 7: GT-time and Group# correlation with globulin concentration and albumin/globulin. Linear regression and regression coefficient.

Dependent variable	Independent variable (equation)	r
GT-time	- 0,22 [globulin] + 16,33	0,61
GT-time	10,84 [albumin/globulin] - 1,21	0,67
Group #	- 0,10 [mean globulin within groups] + 6,50	0,96
Group #	4,06 [mean albumin/globulin within groups] - 0,83	0,96

Among the 37 horses, the proportion of test negatives of horses that were acutely inflamed was 75% (9/12). The proportion of acutely inflamed test negatives was significantly greater than the proportion of chronically inflamed test positives ($p=0.04$, Chi square test). The proportion of test positives of horses that were chronically inflamed was 40% (10/25).

The GT did not show statistically significant correlation with the concentration of blood fibrinogen in acute or chronic diseases.

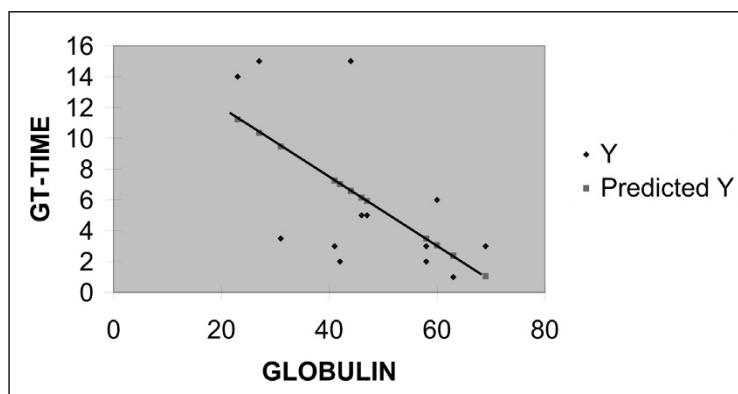
All results from the blood analyses (Table 3) were also compared to the GT using principal component analysis without finding any statistically significant correlation.

Discussion

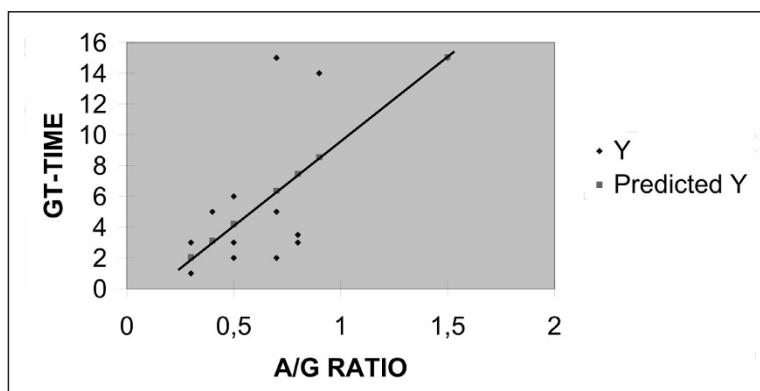
The results of this study indicate that the GT can be used to quickly differentiate chronic from acute inflammatory disease in horses. The

high proportion of test negatives of horses having acute inflammation indicates that horses with inflammatory disease and negative GT are likely to be acutely, rather than chronically, inflamed. Among GT positive horses, 77% were chronically inflamed as shown in Table 5. The GT was not reliable in predicting the blood concentration of fibrinogen in acute or chronic inflammatory diseases.

Useful clinical information could be obtained by dividing GT-times into categories (groups) as listed in Table 2 (*Liberg et al.* 1975a, *Liberg et al.* 1975b). Comparison of category and respectively globulin concentration and albumin/globulin ratio within a category seemed to correlate, although this tendency was not statistically significant. This could be due to the small number of data points. A larger number of horses included in a future study like ours would probably eliminate this statistical uncer-



Graph 1.
GT-time/globulin
regression.

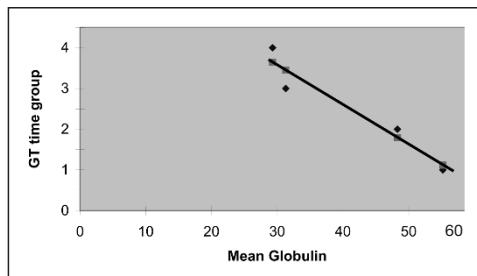


Graph 2.
GT-time/A/G ratio
regression.

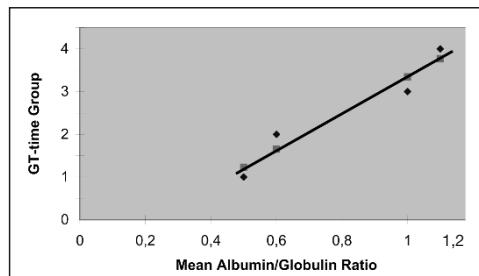
tainty. The correlation above has been observed in cattle (*Sandholm 1974a, Liberg et al. 1975a, Liberg et al. 1975b, Nielsen 1975, Eriksen 1984*). The difference between other studies of other species and this study was that horses in Group 1 had only moderately increased globulin concentration and moderately decreased albumin/globulin ratio, Group 2 horses had a mildly increased globulin concentration and mildly decreased albumin/globulin ratio, and horses in Group 3 had a globulin concentration and albumin/globulin ratio within normal range.

If the clinical examination indicates systemic infection (eg. increased rectal temperature) and the GT is positive, the probability is high (77% likelihood) for chronic inflammatory disease. A

positive GT acts then as an indicator for further laboratory analysis of blood to determine chronicity and etiology of the disease. If the test is negative, the disease is most likely acute or the systemic inflammatory response is either insignificant or absent. The GT can also be used as an additional diagnostic test to indicate prognosis because a positive test predicted fatal outcome in 69% of the clinical cases we studied. The test performance regarding the predictability of a fatal outcome might increase if only severe inflammatory diseases are included as compared to a study also including mild cases (selection bias). Also, the lack of controls will add bias to the percentages and will eliminate false positives. Because the study did not include a group of controls and a group of



Graph 3. Regression of GT time Group vs Mean Globulin.



Graph 4. Regression of GT-time Group vs Mean Albuimin/Globulin Ratio.

horses suffering from non-inflammatory diseases, the data presented can only be considered valid for horses with inflammatory disease. For this reason, the conclusions are not valid for the entire population of horses. The selection of horses among patients submitted to a large referral hospital also might introduce spectrum bias as the hospitalized horses are more likely to be severely affected than horses treated in practice.

A positive GT in horses indicated the probability of increased serum concentration of globulin and a decreased albumin/globulin ratio, but the GT was not correlated with the blood concentration of fibrinogen.

Taking into consideration the low cost and rapid application of the GT and correlation of a positive test with increased concentration of globulin, the GT is a useful screening test for horses suspected to suffer from inflammatory disease.

a) Glutarvac Test tube; Jorgen Kruuse A/S, Marslev, Denmark.

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References

- Aasted B, Leslie G, Agger R: Immunologi. (Immunology). DSR-forlag Landbohøjskolen, Copenhagen 1989, p. 29-36.
- Beetson SA, Hilbert BJ, Mills JN: The use of the glutaraldehyde coagulation test for detection of hypogammaglobulinaemia in neonatal foals. Aust. Vet. J. 1985, 62, 279-281.
- Bendixen HJ: Investigations on the relationship between the serum proteins and the formol-gel reaction in cattle. Nord. Vet.-Med. 1954, 6, 187-194.
- Bruijn CM, Wensing T, Nieuwstadt RA, Bruijn CM, Nieuwstadt RA: Een onderzoek naar de betrouwbaarheid van de glutaraldehydetest voor de bepaling van het gammaglobulinegehalte in het serum van veulens en naar de bruikbaarheid van deze test in de praktijk bij het controleren van de colostrumopname bij veulens. (A study of the reliability of the glutaraldehyde test to determine the level of gamma globulin in the serum of foals and the suitability of this test in practice for the control of colostrum intake in foals). Tijdschr. Diergeneesk. 2003, 128, 240-246.
- Carstairs-Grant SJ, Crawshaw GJ, Mehren KG: A comparison of the glutaraldehyde coagulation test and total serum protein estimation as indicator of gamma globulin levels in neonatal ruminants. J. Zoo. Anim. Med. 1988, 19, 14-17.
- Chadli M, Mahin L: Test d'étable au glutaraldehyde, indicateur préliminaire de la pathologie infectieuse chronique au sein d'une exploitation bovine. (The herd glutaraldehyde test, preliminary indicator of chronic infectious disease in cattle). Act. Int. Agron. Vet. Hassan II. 1986, 6, 49-57.
- Clabough DL, Conboy HS, Roberts MC: Comparison of four screening techniques for the diagnosis of equine neonatal hypogammaglobulinemia. J. Am. Vet. Med. Assoc. 1989, 194, 1717-1720.
- Clabough DL, Levine JF, Grant GL, Conboy HS: Factors associated with failure of passive transfer of colostral antibodies in standardbred foals. J. Vet. Intern. Med. 1991, 5, 335-340.
- Doll K, Schillinger D, Klee W: Der Glutaraldehyd-Test beim Rind-seine Brauchbarkeit für Diagnose und Prognose innerer Entzündungen. (Suitability of the glutaraldehyde test for diagnosis and prognosis of internal inflammatory conditions in cattle). Zentralbl. Vet. Med. 1985, 32, 581-593.
- Eriksen L: Klinisk undersøgelsesmetodik og journalskrivning. (Methods in clinical examination and record writing). CF Mortensen A/S, Copenhagen 1984, p. 87-88+134.
- Ezhilan V, Bhuvanakumar CK: Forecasting of IgG in neonates from colostrum. Centaur Mylap. 1998, 15, 39-40.
- Hansen K: Glutaraldehydpøvens anvendelighed på svin, der ønskes nødslaget. (The applicability of the glutaraldehyde test in slaughter swine). Dansk Vet. Tidsskr. 1985, 68, 151-156.
- Jones D, Brook D: Investigation of the Gamma-Check-C test as a mean of evaluating IgG levels in equine colostrum. J. Equine Vet. Sci. 1995, 15, 269-271.
- Juyal PD, Uppal SK: Determination of gamma globulins in young buffalo calves by glutaraldehyde coagulation test. Indian J. Anim. Health 1995, 34, 161-162.
- Kalinbacak A, Or ME: Yenidogan taylarda hipogam-

- magglobulinemi'nin saptanmasında glutaraldehit koaglyon testi'nin kullanımı. (Use of the glutaraldehyde coagulation test to detect hypogammaglobulinaemia in newborn foals). Vet. Fakult. Dergisi Ankara Univ. 1996, 43, 203-207.
- Kantor IN, Lopez B, Torres P, Nader A, Garcia V, De-Kantor IN:* Preliminary evaluation of a simple method for detection of bovine tuberculosis: the glutaraldehyde test. J. Vet. Med. Series B. 1993, 40, 27-30.
- Katholm J, Jorgensen RJ:* Glutaraldehyd test. Til "cow side" påvisning af colimastitis. (The glutaraldehyde test for "cow side" diagnosis of acute coliform mastitis). Dansk Vet. Tidsskr. 1992, 75, 486-487.
- Keulen KAS, Dobbelaar P, Noordhuizen JPTM, Schwering C, Wensing T:* Een onderzoek naar een aantal aspecten van de biestverstrekking op melkveebedrijven en naar de bruikbaarheid van de glutaraldehydetest bij de beoordeling van de biestverstrekking. (Studies on a number of features of the supply of colostrum on dairy farms and the use of the glutaraldehyde test in evaluating the supply of colostrum). Tijdschr. Diergeesk. 1984, 109, 605-611.
- Kovac G:* Diagnosis of hypogammaglobulinemia in calves by means of the glutaraldehyde coagulation test. Folia Vet. 1988, 32, 71-78.
- Kovac G, Bartko P, Mudron P, Michna A, Bires J, Bal-dovic J:* Glutaraldehydovy koagulacny test u osipanych. (Glutaraldehyde coagulation test in pigs) Sloven. Vet. Casop. 1993, 18, 66-68.
- Kumaran D, Bhuvanakumar CK:* Detection of immunoglobulin levels in neonatal foals. Centaur Mylap. 1994, 10, 98-100.
- Larsson B:* The relationship between total protein in serum, glutaraldehyde coagulation test and disease in feedlot calves. Nord. Vet.-Med. 1985, 37, 90-96.
- Liberg P, Pehrson B, Sandholm M:* Snabbstest för diagnostisering av inflammatoriska tilstånd hos nötkreatur. (Rapid test for diagnosis of inflammatory disease in cattle). Svensk Vet. Tidn. 1975a, 27, 181-183.
- Liberg P, Pehrson B, Sandholm M:* The value of the glutaraldehyde and formaldehyde tests in evaluation of the globulin level in bovine blood. Acta Vet. Scand. 1975b, 16, 236-243.
- Liberg P:* The fibrinogen concentration in blood of dairy cows and its influence on the interpretation of the glutaraldehyde and formol-gel test reactions. Acta Vet. Scand. 1978, 19, 413-421.
- Liberg P:* Helblodstestning med glutaraldehyd vid svinslakt - en preliminär undersökning. (Glutaraldehyde test on whole blood of slaughter swine). Nord. Vet.-Med. 1979, 31, 360-366.
- Liberg P:* Glutaraldehyde and formol-gel tests in bovine traumatic peritonitis. Acta Vet. Scand. 1981, 22, 78-84.
- Liberg P:* Blood protein screening in healthy and diseased cattle. Agarose gel electrophoresis, the formol-gel and glutaraldehyde tests. The Swedish Veterinary University, Uppsala 1982.
- Mahlin L, Chadli M, Marzou A, Maach L, Ychou M:* Differences in coagulability of three glutaraldehyde solutions in the glutaraldehyde test on bovine whole blood. Zentbl. Vet. Med. 1985, 32, 151-154.
- Martens HH:* Untersuchungen mit der Glutaraldehydprobe nach Sandholm im Vollblut gesunder und kranker Rinder. (Application of Sandholm's glutaraldehyde test to whole blood from healthy and diseased cattle). Thesis, Hannover 1977.
- Martin DWJ, Mayes PA, Rodwell VW, Granner DK:* Harper's review of biochemistry. 20th ed. Lange Medical Publications, California 1985, p. 637-645.
- Nansen P, Nielsen K:* Metabolism of bovine immunoglobulin. 1. Metabolism of bovine IgG in cattle with chronic pyogenic infections. Can. J. Comp. Med. Vet. Sci. 1966, 30, 327-331.
- Nielsen K:* Glutaraldehydproven, en metode til påvisning af forhøjet immunoglobulin koncentration i blod. (The glutaraldehyde test, a method for determination of increased concentration of immunoglobulin in blood). Dansk Vet. Tidsskr. 1975, 58, 652-655.
- O'Rourke KI, Satterfield WC:* Glutaraldehyde coagulation for detection of hypogammaglobulinemia in neonatal nondomestic ruminants. J. Am. Vet. Med. Assoc. 1981, 179, 1144-1146.
- Ramprabhu R, Dhanapalan P, Prathaban S:* Efficacy of Sulkowitch and glutaraldehyde tests in traumatic reticuloperitonitis and allied syndrome in cattle. Indian J. Anim. Health 2002, 41, 74-76.
- Saikku A, Koskinen E, Sandholm M:* Detection of hypogammaglobulinaemia in neonatal foals using the glutaraldehyde coagulation test. J. Vet. Med. Series B. 1989, 36, 168-174.
- Sandholm M, Kangas J:* Coagulation of hyperglobulinaemic mink blood by glutaraldehyde. Zentbl. Vet. Med. 1973, 20B, 206-211.
- Sandholm M:* A preliminary report of a rapid method for the demonstration of abnormal gammaglobu-

- lin levels in bovine whole blood. *Res. Vet. Sci.* 1974a, *17*, 32-35.
- Sandholm M:* Die Feststellung der Hyper- α -Globulinämie beim Rind unter Praxisbedingungen. (The determination of hyperglobulinemia in cattle under practice conditions). *Tierärztl. Prax.* 1974b, *2*, 237-240.
- Sandholm M, Kivistö AK:* Determination of gamma globulin in dog serum by glutaraldehyde. *J. Small. Anim. Pract.* 1975, *16*, 201-205.
- Satpathy PK, Dutta NK, Mishra PR, Kar BC:* Glutaraldehyde coagulation test: standard curve and its applications to detect gammaglobulin level in kids. *Indian Vet. J.* 1996, *73*, 257-260.
- Sen I, Basoglu A, Ok M, Birdane FM, Guzelbektas H, Civelek T:* Neonatal ishalli buzagılarda serum immunoglobulinlerin glutaraldehid koagulasyon testi ile degerlendirilmesi. (Serum immunoglobulins in neonatal diarrhoeic calves evaluation by glutaraldehyde coagulation test). *Vet. Bilim. Dergisi* 2000, *16*, 143-146.
- Tennant B, Baldwin BH, Braun RK, Norcross NL, Sandholm M:* Use of the glutaraldehyde coagulation test for detection of hypogammaglobulinemia in neonatal calves. *J. Am. Vet. Med. Assoc.* 1979, *174*, 848-853.
- Tyler JW, Besser TE, Wilson L, Hancock DD, Sanders S, Rea DE:* Evaluation of a whole blood glutaraldehyde coagulation test for the detection of failure of passive transfer in calves. *J. Vet. Intern. Med.* 1996, *10*, 82-84.
- Vihan VS:* Glutaraldehyde coagulation test for detection of hypo-gammaglobulinaemia in neonatal kids. *Indian Vet. J.* 1989, *66*, 101-105.
- Wolff B:* Test à la glutaraldehyde: une méthode d'appoint dans le diagnostic du pyometre chez la chienne. (Glutaraldehyde test: a supplementary diagnostic method for pyometra in the bitch). *Point Vet.* 1986, *18*, 69-71.

Sammendrag

Glutaraldehydprøvens evne til at skelne mellem akut og kronisk inflammatorisk sygdom hos hest.

Glutaraldehydprøven (GP), en hurtig og billig test, har været anvendt empirisk gennem mange år i kvægpraksis for diagnosticerig af inflammatoriske sygdomme. GP bliver primært brugt til at påvise øget serum koncentration af fibrinogen og globulin. Glutaraldehyd bindes til frie amino-grupper i fibrinogen og globulin, som derpå danner et blodkoagel ved en 1. grads kemisk reaktion. Koaguleringsiden af GP estimerer indholdet af de proteiner, som produceres i et inflammatorisk respons. Anvendeligheden af GP til diagnosticerig af inflammatoriske tilstande i hestepraksis har aldrig været undersøgt før. Formålet med dette studie er at bestemme GPs evne til at skelne mellem akut og kronisk inflammatorisk sygdom hos hest. 37 heste, mistænkt for inflammatorisk sygdom, blev evalueret på basis af GP, anamnese, fuldstændig klinisk undersøgelse samt rutinemæssig blodprøver. GP-tid, blodprøvesvar og klinisk udfald blev sammenlignet statistisk. De heste, som var bestemt til at være akut afficeret på basis af anamnese, klinisk undersøgelse og rutinemæssig blodprøve, tenderede mod at have negativ GP (75%). Der kunne ikke påvises sammenhæng mellem GP og fibrinogen koncentration i blodet. Positiv GP forudsagde også et fatalt udfald i 69% af de kliniske tilfælde. Resultaterne af dette studie indikerer, at GP kan være en brugbar praktisk test til at skelne mellem akut og kronisk inflammatorisk sygdom hos hest.

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