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Effect of sedation with detomidine and butorphanol on pulmonary gas exchange in the horse

Görel Nyman*¹, Stina Marntell², Anna Edner³, Pia Funkquist³, Karin Morgan⁴ and Göran Hedenstierna⁵

Address: ¹Department of Environment and Health, Faculty of Veterinary Medicine and Animal Science, Swedish University of Agricultural Sciences, Skara, Sweden, ²Orion Pharma Animal Health, Sollentuna, Sweden, ³Department of Clinical Sciences, Faculty of Veterinary Medicine and Animal Science, Swedish University of Agricultural Sciences, Uppsala, Sweden, ⁴Department of Equine Studies, Faculty of Veterinary Medicine and Animal Science, Swedish University of Agricultural Sciences, Uppsala, Sweden and ⁵Department of Medical Sciences, Clinical Physiology, University Hospital, Uppsala, Sweden

E-mail: Görel Nyman* - gorel.nyman@gmail.com; Stina Marntell - stina.marntell@orionpharma.com; Anna Edner - anna.edner@kv.slu.se; Pia Funkquist - pia.funkquist@kalmar.nshorse.se; Karin Morgan - karin.morgan@stromsholm.com; Göran Hedenstierna - goran.hedenstierna@medsci.uu.se

*Corresponding author

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Abstract

Background: Sedation with α_2 -agonists in the horse is reported to be accompanied by impairment of arterial oxygenation. The present study was undertaken to investigate pulmonary gas exchange using the Multiple Inert Gas Elimination Technique (MIGET), during sedation with the α_2 -agonist detomidine alone and in combination with the opioid butorphanol.

Methods: Seven Standardbred trotter horses aged 3–7 years and weighing 380–520 kg, were studied. The protocol consisted of three consecutive measurements; in the unsedated horse, after intravenous administration of detomidine (0.02 mg/kg) and after subsequent butorphanol administration (0.025 mg/kg). Pulmonary function and haemodynamic effects were investigated. The distribution of ventilation-perfusion ratios (V_A/Q) was estimated with MIGET.

Results: During detomidine sedation, arterial oxygen tension (PaO_2) decreased (12.8 ± 0.7 to 10.8 ± 1.2 kPa) and arterial carbon dioxide tension ($PaCO_2$) increased (5.9 ± 0.3 to 6.1 ± 0.2 kPa) compared to measurements in the unsedated horse. Mismatch between ventilation and perfusion in the lungs was evident, but no increase in intrapulmonary shunt could be detected. Respiratory rate and minute ventilation did not change. Heart rate and cardiac output decreased, while pulmonary and systemic blood pressure and vascular resistance increased. Addition of butorphanol resulted in a significant decrease in ventilation and increase in $PaCO_2$. Alveolar-arterial oxygen content difference $P(A-a)O_2$ remained impaired after butorphanol administration, the V_A/Q distribution improved as the decreased ventilation and persistent low blood flow was well matched. Also after subsequent butorphanol no increase in intrapulmonary shunt was evident.

Conclusion: The results of the present study suggest that both pulmonary and cardiovascular factors contribute to the impaired pulmonary gas exchange during detomidine and butorphanol sedation in the horse.

Background

The possibility of producing potent sedation of horses by alpha-2-adrenoreceptor agonists (α_2 -agonists) is one of the greatest improvements in modern equine practice. The dose-dependent sedation and analgesia produced by the α_2 -agonists is reliable for diagnostic procedures and for treatment of various conditions. The central action of the α_2 -agonist is a presynaptic inhibition of noradrenaline accompanied by a decreased sympathetic tone [1]. Alpha-2-agonists also exert physiological effects by their action on peripheral α_2 -receptors [2]. Besides the well recognised and potent cardiovascular changes, sedation with α_2 -agonists in the horse is reported to be accompanied by impairment of pulmonary gas exchange and arterial oxygenation [3-6]. From the studies reported in the horse to date, it is not possible to separate the relative contributions of pulmonary and cardiovascular alterations to the development of impaired arterial oxygenation.

Horses that are deeply sedated with an α_2 -agonist are not unconscious. A sedated horse must be handled with caution, since it may be aroused by stimulation and can respond with dangerous kicks [7-9]. In a situation in which a painful procedure is planned or local analgesia needs to be placed before surgery on the standing horse, accentuation of both sedation and analgesia can be achieved by adding an opioid to the α_2 -agonist [4,10,11]. Butorphanol, a mixed opioid with agonistic and antagonistic properties, has proven effective in such a combination [3,4,12]. There are limited reports on the respiratory effects of butorphanol alone or in combination with the α_2 -agonist detomidine in horses [5,11], but the effects of the combination on pulmonary gas exchange has not been clarified.

With the multiple inert gas elimination technique, developed by Wagner et al. [13] and modified for use in the standing horse [14], the pulmonary gas exchange and a virtually continuous distribution of ventilation-perfusion ratios can be studied.

The aim of the present investigation was to determine the physiological effects, especially on the pulmonary gas exchange, of sedation with detomidine alone and in combination with butorphanol.

Methods

Horses

Seven Standardbred trotters (two mares and five geldings) that were considered healthy on clinical examination were studied. Their mean weight was 457 kg (range 380–520 kg) and mean age 5 years (range 3–7 years). Food and water were withheld for approximately 3 hours

prior to the sedation procedure. The local Ethical Committee on Animal Experimental in Uppsala, Sweden approved the experimental procedure.

Catheterisation

All catheterisations were performed with the horse standing and unsedated, after local analgesia with lidocaine (Xylocain® 2%, Astra, Sweden). A catheter was introduced percutaneously into the transversal facial artery (18G, Hydrocath TM arterial catheter, Omeda, UK) for systemic arterial blood pressure measurements and collection of arterial blood. A 100 cm pigtail catheter (Cook Europe A/S, Söborg, Denmark) for injection of ice cold saline during thermodilution measurements was introduced by the same technique into the right jugular vein, advanced to the right ventricle and then retracted into the right atrium under pressure-tracing guidance. A thermodilution catheter (7F, Swan-Ganz, Edwards lab., Santa Ana, CA, USA) was inserted with an introducer kit (8F, Arrow Int. Inc., Reading, PA, USA) into the right jugular vein and advanced into the pulmonary artery for mixed venous blood sampling and measurements of core temperature and pulmonary arterial blood pressure. Once correctly placed, the catheters were locked in position with Luer-lock adapters. Further, two infusion catheters (14G, Intranule, Vygon, France) were placed in the left jugular vein.

Protocol

Detomidine 0.02 mg/kg (Domosedan® vet., 10 mg/ml, Orion Pharma Animal Health, Sollentuna, Sweden) was given intravenously (IV), followed 20 minutes later by butorphanol 0.025 mg/kg IV (Torbugesic®, 10 mg/ml, Fort Dodge Animal Health, Fort Dodge, IA, USA). Sampling of blood and expired gas for measurements of gas concentrations by the multiple inert gas elimination technique (MIGET) were performed in the unsedated standing horse (Unsedated) and started 15 minutes after the detomidine injection (Detomidine) and 15 minutes after the butorphanol injection (Detomidine + Butorphanol). The order of the measurements was the same on each occasion, haemodynamic parameters followed by pulmonary function and gas exchange, and the sampling was completed in 5 minutes.

Measurements of haemodynamic parameters

Systemic arterial and pulmonary arterial blood pressure (SAP and PAP) were measured by connecting the arterial catheters via fluid-filled lines to calibrated pressure transducers (Baxter Medical AB, Eskilstuna, Sweden) positioned at the level of the scapulo-humeral joint. Blood pressure and electrocardiogram (ECG) were recorded on an ink-jet recorder (Sirecust 730, Siemens-Elema, Solna, Sweden). Heart rate (HR) was recorded

from the ECG. Cardiac output (Qt) was determined by the thermodilution technique (Cardiac Output Computer Model 9520A, Edwards lab., Santa Ana, CA, USA). A bolus of 20 ml ice cold 0.9% saline was rapidly injected into the right atrium through the pigtail catheter (injection time 3 sec), and the blood temperature was then measured in the pulmonary artery at the tip of the Swan-Ganz catheter and the cardiac output was computed from the recorded temperature change. The mean of at least three consecutive measurements was used.

Measurements of pulmonary function and gas exchange

Respiratory rate (RR) was measured by observing the costo-abdominal movements, and expired minute ventilation (V_E) was measured with a Tissot spirometer, range 0.5–685 l (Collins inc., Braintree, MA, USA) attached to the nose mask. Oxygen uptake (VO_2) was determined by analysing gas from mixed expired air with a calibrated gas analyser (Servomex, Sussex, UK, integrated into an Oximeter 3200, Isler Bioengineering AG, Switzerland). Volume and gas parameters are measured at body temperature and pressure saturated (BTPS). Arterial (a) and mixed venous (v) blood samples for measurements of oxygen and carbon dioxide tensions (PaO_2 , PvO_2 , $PaCO_2$, $PvCO_2$) and oxygen saturation of haemoglobin (SaO_2 , SvO_2) were drawn simultaneously and anaerobically into heparinised syringes and stored on ice until analysed (within 30 minutes) by means of conventional electrode techniques with correction of the $p50$ value (ABL 300 and Hemoximeter OSM 3, Radiometer, Copenhagen, Denmark). Haemoglobin concentration [Hb] was determined spectrophotometrically (Ultralab system, 2074 Calculating Absorptiometer LKB Clinicon, Bromma, Sweden).

The distribution of ventilation and perfusion was estimated by the multiple inert gas elimination technique [13,14]. Six gases (sulphur hexafluoride, ethane, cyclopropane, enflurane, diethyl ether and acetone), inert in the sense of being chemically inactive in blood, were dissolved in isotonic Ringer acetate solution (Pharmacia, Stockholm, Sweden) and infused continuously into the jugular vein at 30 ml/min from at least 40 minutes before baseline measurements until the collection of the last samples, 15 minutes after butorphanol injection. Arterial and mixed venous blood samples were drawn and simultaneously mixed expired gas was collected from a heated mixing box connected to a nose mask. Gas concentrations in the blood samples and expirate were measured by the method of Wagner et al. [15], using a gas chromatograph (Hewlett Packard 5890 series II, Atlanta, GA, USA). The arterial/mixed venous and mixed expired/mixed venous concentration ratios of each gas (retention and excretion, respectively) depend

on its blood-gas partition coefficient and the V_A/Q (the ratio of alveolar ventilation, V_A and cardiac output, Q) of the lung. The retention and excretion were calculated for each gas, and the solubility of each gas in blood was measured in each horse by a two-step procedure [15]. The solubilities were similar to those reported previously [14]. These data were then used for deriving the distribution of ventilation and blood flow in a 50-compartment lung model, with each compartment having a specific alveolar ventilation/blood flow ratio (V_A/Q ratio) ranging from zero to infinity. Ventilation and blood flow in healthy subjects have a log normal distribution against V_A/Q ratios. Of the information obtained concerning the V_A/Q distribution, data are presented for the mean and standard deviation of the blood flow log distribution (Q_{mean} and log SDQ, respectively), shunt (perfusion of lung regions with $V_A/Q < 0.005$), and the mean and standard deviation of the ventilation log distribution (V_{mean} and log SDV, respectively). All subdivisions of blood flow and ventilation are expressed in per cent of cardiac output and expired minute ventilation, respectively. The difference between measured PaO_2 and PaO_2 predicted from MIGET-algorithms on the basis of the amount of ventilation-perfusion mismatching and shunt was determined. A higher predicted than measured PaO_2 may reflect diffusion limitation or extrapulmonary shunt.

Calculations and statistics

From the measurements obtained the following calculations were made, using standard equations. Stroke volume (SV), systemic vascular resistance (SVR) and pulmonary vascular resistance (PVR) as follows:

$$SV = Qt / HR$$

$$SVR = \text{mean SAP} / Qt$$

$$PVR = (\text{mean PAP} - \text{diastolic PAP}) / Qt$$

Diastolic PAP was used in the formula as a substitute for wedge pressure.

For the following calculations, blood gas values measured at 37°C were used.

Alveolar oxygen partial pressure: $PAO_2 = (P_1O_2 - (PaCO_2/R))$

($R =$ Respiratory exchange ratio = 0.8), where $P_1O_2 =$ partial pressure of inspired O_2 .

The alveolar – arterial oxygen tension difference ($P(A-a)O_2$) was calculated.

Content of oxygen in arterial (a), mixed venous (v), and end-capillary pulmonary (c) blood:

$CzO_2 = (\text{Hb concentration} \times 1.39 \times \text{oxygen saturation of Hb}) + (PzO_2 \times 0.003)$, where $z = a, v, c$. $PcO_2 \approx PaO_2$.

Arterial-mixed venous oxygen content difference ($C(a-v)O_2$) = $CaO_2 - CvO_2$.

Oxygen delivery: $O_2\text{-del} = CaO_2 \times Qt$.

Cardiac output (Qt) was also computed through mass balance from measured VO_2 and the arterio-venous oxygen (or inert gas) content difference (the Fick principle). The cardiac output measurements presented in Table three are based on thermodilution measurements.

For statistical analysis the Statistica 6.0 software package (Statsoft Inc., Tulsa, OK, USA) was used. The data were analysed in a General Linear Model with repeated measures

ANOVA. When the ANOVA indicated a significant difference, Tukey's HSD post hoc test was used to determine at what time point there were significant differences within the protocol from baseline and sedation, unless Mauchly's sphericity test indicated significance. In this instance, a planned comparison was applied to define the contrast at each treatment [16]. A p-value less than 0.05 was considered significant. Results are given as mean values \pm SD.

Results

Data on ventilation and blood gases are presented in Table 1, pulmonary gas exchange based on inert gas data in Table 2 and circulation in Table 3.

Unsedated horse

In the unsedated, standing horse, circulatory data as well as ventilation and pulmonary gas exchange (Tables 1, 2 and 3) were all within normal limits [14]. The distribution of ventilation and perfusion was centered upon a V_A/Q ratio of approximately 1 ($Q_{\text{mean}} = 0.79$) in all horses (Figure 1,

Table 1: Circulatory data (n = 7)

		Unsedated	Detomidine	Detomidine-butorphanol	GLM – ANOVA
HR	Beats/min	38 \pm 8	23 \pm 5*	29 \pm 5*	p < 0.001
Qt	ml/min \times kg	72 \pm 14	32 \pm 10*	44 \pm 6*	p < 0.001
SV	ml/kg \times beat	2.0 \pm 0.7	1.4 \pm 0.6	1.5 \pm 0.3	NS
SAP mean	mmHg	116 \pm 15	148 \pm 14*	137 \pm 14*	p < 0.001
PAP mean	mmHg	26 \pm 2	34 \pm 3*	31 \pm 4*	p < 0.001
SVR	mmHg/ml/min \times kg	1.69 \pm 0.49	5.01 \pm 1.45*	3.16 \pm 0.62 *†	p < 0.001
PVR	mmHg/ml/min \times kg	0.15 \pm 0.06	0.31 \pm 0.16*	0.15 \pm 0.06†	p = 0.017
O₂ del	ml/min \times kg	11.4 \pm 2.6	5.1 \pm 1.8*	6.5 \pm 0.8*	p < 0.001
C(a-v)O₂	ml/100 ml	6.1 \pm 0.8	8.5 \pm 1.8*	7.3 \pm 1.1	p = 0.002
Hb	g/l	1.15 \pm 1.0	1.18 \pm 1.3	1.11 \pm 1.2	NS (p = 0.062)

Data presented as mean \pm SD for heart rate (HR), cardiac output thermodilution (Qt), stroke volume (SV), mean systemic arterial pressure (SAP mean), mean pulmonary arterial pressure (PAP mean), systemic vascular resistance (SVR), pulmonary vascular resistance (PVR), oxygen delivery (O_2 del), arterial-mixed venous oxygen content difference ($C(a-v)O_2$) and haemoglobin concentration (Hb). Results for General Linear Model-ANOVA (GLM-ANOVA), p value for differences between the treatments, NS = non-significant. Differences between treatments are presented with the abbreviations: * = significantly different from unsedated, † = significantly different from detomidine sedation.

Table 2: Ventilation and blood gases (n = 7)

		Unsedated	Detomidine	Detomidine-butorphanol	GLM – ANOVA
RR	Breaths/min	16 \pm 3	12 \pm 5	10 \pm 1* †	p = 0.032
V_E	ml/min \times kg	163 \pm 36	157 \pm 42	114 \pm 24*	p = 0.031
VT	ml/kg	8.6 \pm 1.8	10.6 \pm 3.5	10.1 \pm 2.4	NS
PaCO₂	kPa	5.9 \pm 0.3	6.1 \pm 0.2*	6.4 \pm 0.3* †	p < 0.001
	(mmHg)	(44.3 \pm 2.2)	(46.1 \pm 1.8)	(47.7 \pm 2.1)	
P(A-a)O₂	kPa	0.5 \pm 0.4	2.2 \pm 0.7*	2.3 \pm 1.3*	p < 0.001
	(mmHg)	(4.1 \pm 2.8)	(16.6 \pm 5.4)	(17.2 \pm 9.7)	
PaO₂	kPa	12.8 \pm 0.7	10.8 \pm 1.2*	10.6 \pm 1.4*	p < 0.001
	(mmHg)	(95.7 \pm 4.5)	(80.7 \pm 8.7)	(79.2 \pm 10.6)	
PvO₂	kPa	4.3 \pm 0.3	3.5 \pm 0.5*	3.6 \pm 0.2*	p < 0.001
	(mmHg)	(32.5 \pm 2.6)	(26.0 \pm 3.5)	(27.0 \pm 1.7)	
VO₂	ml/min \times kg	3.2 \pm 0.5	2.4 \pm 0.6	2.9 \pm 1.0	NS

Data presented as mean \pm SD for respiratory rate (RR), expired minute ventilation (V_E), tidal volume (VT), arterial carbon dioxide tension ($PaCO_2$), alveolar-arterial oxygen tension difference ($P(A-a)O_2$), arterial oxygen tension (PaO_2), mixed venous oxygen tension (PvO_2) and oxygen uptake (VO_2). For other explanations see Table 1.

Table 3: Ventilation/perfusion relationship (V_A/Q) data (n = 7)

		Unsedated	Detomidine	Detomidine-butorphanol	GLM – ANOVA
Percentage perfusion of regions with:	Shunt	1.1 ± 0.3	1.3 ± 0.4	1.1 ± 0.4	NS
	Normal V_A/Q	98.8 ± 0.4	98.5 ± 0.6	98.8 ± 0.4	NS
Percentage ventilation of regions with:	Normal V_A/Q	36.0 ± 5.2	30.8 ± 7.5	35.7 ± 7.1	NS
	High V_A/Q	0.3 ± 0.5	2.5 ± 3.8	1.6 ± 2.3	NS
	Dead space	63.6 ± 5.1	66.5 ± 4.2	61.3 ± 4.4 †	p = 0.047
	Log SDQ	0.37 ± 0.09	0.45 ± 0.11*	0.41 ± 0.09	p = 0.002
	Log SDV	0.55 ± 0.32	0.85 ± 0.64	0.80 ± 0.59	NS
	Mean Q	0.79 ± 0.21	1.58 ± 0.32*	0.86 ± 0.18 †	p < 0.001
	Mean V	0.95 ± 0.16	2.8 ± 1.7*	1.2 ± 0.33 †	p = 0.029

Data presented as mean ± SD. Log SDQ = logarithmic standard deviation of blood flow (Q) around Q mean (unit V_A/Q ratio.); Shunt = $V_A/Q < 0.005$; normal $V_A/Q = 0.1 < V_A/Q < 10$. Log SDV = logarithmic standard deviation of ventilation (V) around V mean (unit V_A/Q ratio); normal $V_A/Q = 0.1 < V_A/Q < 10$; high $V_A/Q = 10 < V_A/Q < 100$; dead space = (inert gas) including apparatus dead space: $V_A/Q > 100$. For other explanations see Table 1.

top panel). The overall log SDQ, was 0.37. No regions of low V_A/Q were noted and in no case was the shunt larger than 1.5% of cardiac output (Figure 1, top panel). The overall log SDV was 0.55, centered around $V_{mean} = 0.95$. Bimodal ventilation distribution with an additional mode located within high V_A/Q ratios ($V_A/Q > 10$) was seen in two of seven horses. Dead space ($V_A/Q > 100$) (including apparatus dead space, i.e. face mask and non-rebreathing valves of approximately 1 litre) averaged 64%.

Detomidine sedation

Fifteen minutes after detomidine administration, respiratory rate and expired minute ventilation had not changed significantly, but $PaCO_2$ increased slightly but significantly compared to the values in the unsedated horse (Table 1). $P(A-a)O_2$ increased and PaO_2 and PvO_2 decreased during sedation (Table 1). The shunt remained small but the scatter of V_A/Q ratio increased as evidenced by a higher log SDQ. The centre of the distribution of ventilation and perfusion increased and Q_{mean} and V_{mean} were significantly higher than in the unsedated horse (Figure 1, middle panel). Regions with high V_A/Q ratios were observed in three horses. The predicted PaO_2 compared to the measured PaO_2 was slightly but significantly higher compared to values in the standing horse. HR and Qt decreased while increases in vascular resistance and mean SAP and PAP were noted during sedation with detomidine. Second-degree atrio-ventricular (AV) block was recorded during sedation in six of seven horses. VO_2 did not change, but oxygen delivery decreased significantly and $C(a-v)O_2$ was higher during detomidine sedation compared to the values in the unsedated horse.

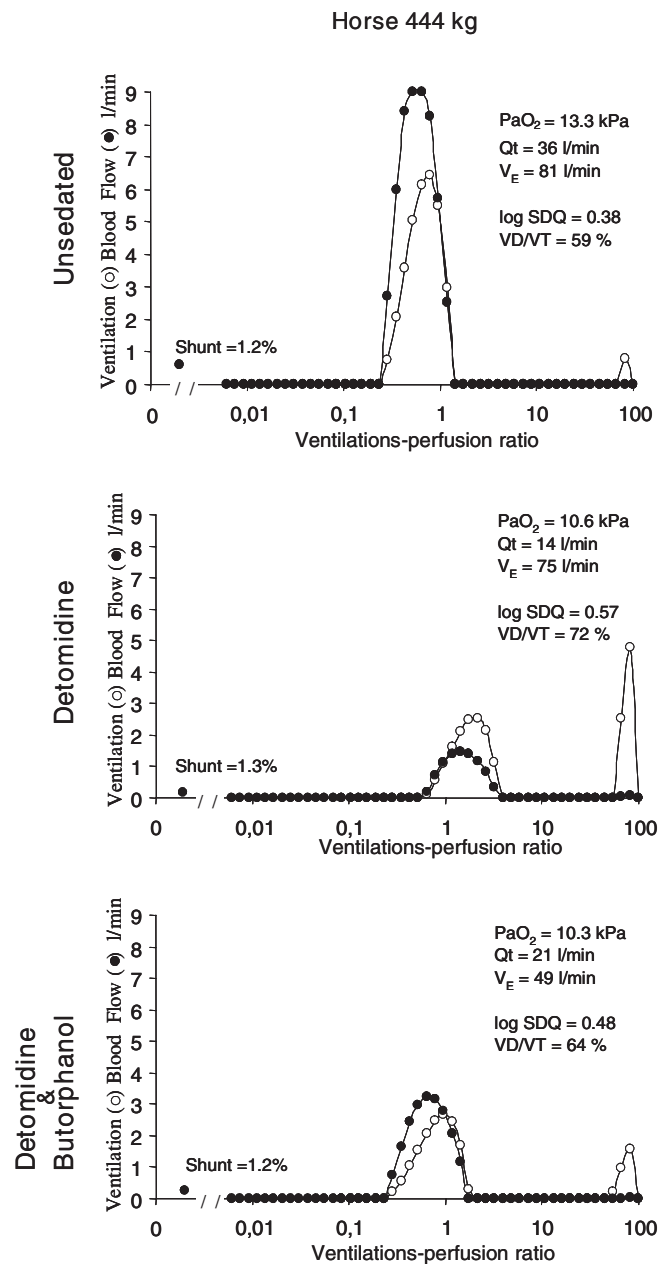
Detomidine and butorphanol combination

Addition of butorphanol during the detomidine sedation resulted in a significant decrease in respiratory rate,

and a small but significant increase in $PaCO_2$ was measured compared to that during detomidine sedation alone (Table 1). Minute ventilation decreased significantly compared to that in the unsedated horse. The cardiovascular changes persisted but the vascular resistance in both the pulmonary and the systemic circulation decreased compared to detomidine sedation alone. Ventilation-perfusion distribution improved and dead space ventilation decreased compared to detomidine sedation. No shunt was seen and the predicted and measured PaO_2 were similar. Q_{mean} and V_{mean} did no longer differ from the unsedated horse (Figure 1, bottom panel). The alterations in $P(A-a)O_2$, PaO_2 and PvO_2 , as well as HR, Qt and mean SAP and PAP, that developed during detomidine sedation remained after addition of butorphanol (Tables 1 and 3). Second-degree AV block remained in five of the six horses which showed AV block during detomidine sedation. $C(a-v)O_2$ decreased and did not longer differ from the unsedated situation.

Discussion

It is suggested in the present study that the impaired pulmonary gas exchange during detomidine and butorphanol sedation in the horse originates from both pulmonary and cardiovascular factors. These results are influenced by time and the order of drug administration since the complexity of performing MIGET, including several physiological measurements, limits the frequency of sampling. In the present investigation first MIGET measurements during sedation was taken 15 minutes after detomidine administration and subsequent MIGET measurements during detomidine and butorphanol sedation were taken 35 minutes after detomidine administration. The most pronounced decrease in heart rate during detomidine sedation has been reported between 2–5 minutes after intravenous administration

**Figure 1**

Distribution of ventilation-perfusion ratio (V_A/Q) in one horse (444 kg). The top panel represent the V_A/Q distribution in an unsedated horse (Unsedated). The middle panel represent the V_A/Q distribution 15 minutes after intravenous detomidine administration (Detomidine). The lower panel represent the V_A/Q distribution 15 minutes after additional intravenous injection of butorphanol (Detomidine & Butorphanol). Note the impaired arterial oxygen tension (PaO₂) during sedation in the middle and bottom panel. During sedation with detomidine, cardiac output (Qt) decreased and there was an increase in ventilation-perfusion mismatch (broader base of ventilation-perfusion ratio and increased SD of blood flow log distribution (log SDQ)) compared to the unsedated horse. The intrapulmonary shunt was minimal. During sedation with detomidine and butorphanol, the impaired PaO₂ was a result of persistent low cardiac output and an additional reduction in expired minute ventilation (V_E). After addition of butorphanol the distribution of V_A/Q improved as the reduced ventilation and persistent low blood flow matched well. No increase in intrapulmonary shunt was evident during subsequent butorphanol administration.

and heart rate remained unchanged between 10 to 30 minutes after injection [8]. In Wagner et al. 1991 [17] detomidine 0.02 mg/kg given intravenously resulted in a significant but stable decrease in cardiac output and respiratory rate compared to unsedated horses between 15 and 60 minutes after administration. In the reported study by Wagner et al. 1991 [17], arterial oxygenation was only significantly decreased at 5 and 15 minutes after sedation. Systemic and pulmonary vascular resistance started to diminish around 30–45 minutes after detomidine injection. The measurements at 15 and 35 minutes after detomidine administration in the present study are thus made at a fairly stable heart rate and cardiac output conditions. The effects on pulmonary gas exchange and oxygenation measured at 35 minutes after sedation is most likely an effect of the additional administration of butorphanol.

Unsedated horse

The good match between ventilation and perfusion in the standing unsedated horse results in near optimal oxygenation. The narrow distribution of perfusion, with absence of low V_A/Q regions, negligible intrapulmonary shunt and no diffusion limitation of oxygen, were similar to that found in previous studies [14,18]. The presence of a high V_A/Q mode, which is usually seen in the resting horse [14], was noted in two of the horses. Interestingly, the horse is able to match ventilation and perfusion as efficiently as young human adults [19,20] and better than sheep [21] despite the fact that the horse has a high vertical lung distance gradient. This shows that the mechanisms for matching ventilation and perfusion are highly efficient in the athletic horse. These mechanisms are probably related to the lung structure and it is proposed that the horse primarily depends for the matching on hypoxic vasoconstriction, i.e. redistribution of blood flow from regions of low ventilation to areas of higher ventilation, by pulmonary vasoconstriction, with only a small contribution from collateral ventilation [22]. Regional PVR is higher in dependent lung regions than in upper ones in the standing horse [23] and this may contribute to the good V_A/Q match.

Detomidine sedation

The impaired pulmonary gas exchange and arterial oxygenation during detomidine sedation in the present study reconfirm previous observations during sedation of horses with α_2 -agonists [3,17,24]. Although the reportedly classic causes of an increased $P(A-a)O_2$, namely ventilation-perfusion mismatch, failure of alveolar-end capillary diffusion equilibration and right-to-left vascular shunt, have been proposed as presumable mechanisms, extrapulmonary contributors, e.g. extrapulmonary shunt and cardiac output alterations, are possible [17].

It has been reported that the physiological changes induced by α_2 -agonist may be dose-dependent [17,25]. Also, since the physiological effects induced by α_2 -agonists are transient, the choice of methodology and time points for data sampling probably affect the results. The detomidine dose of 0.02 mg/kg used in the present study is a clinically effective sedative dose in most horses [8]. The measurements of cardiovascular and pulmonary function were performed at 15 minutes after intravenous injection of the detomidine. The significant increase in $P(A-a)O_2$ was mainly attributed to increased V_A/Q mismatch as a reduction of cardiac output.

The cardiac output was reduced by 56% which is in line with the literature [17,26]. Since, the cardiac output measurement may be inaccurate during bradycardia with AV block, cardiac output was both measured by thermodilution and calculated according to the Fick principle. The results were in good agreement. In the present study no increase in either pulmonary shunt or low V_A/Q was evident in the horses (Figure 1). The significantly increased V_A/Q mismatch (log SDQ) measured during sedation might be caused by a larger vertical difference in perfusion. The shift of the V_A/Q distribution to a higher range of V_A/Q ratios during detomidine sedation (Figure 1) was caused by a significant reduction in pulmonary perfusion with unaltered ventilation.

In the healthy human or animal the expected response on increased V_A/Q mismatch is mitigated by an increase in the overall lung V_A/Q ratio, thereby increasing the alveolar ventilation and raising both alveolar and arterial PO_2 [17,26,27]. The absence of ventilatory response to the detomidine-induced hypoxaemia may be due either to decreased ventilatory responsiveness or to decreased receptor sensitivity. However, in the present study, detomidine administration did not result in changes in respiratory rate or minute ventilation. An unaffected respiratory rate is in line with some reports, although others have found a decreased or increased respiratory rate in healthy detomidine-sedated horses [24,28].

Interestingly, Wagner et al. [17] reported that the respiratory rate was significantly reduced 15 minutes after sedation and remained low during the study period of two hours. Also, the slightly increased $PaCO_2$ suggested that there was some degree of hypoventilation. The lack of a compensatory increase in alveolar ventilation during sedation with α_2 -agonists means that the arterial blood gases are not corrected. It has been demonstrated that α_2 -adrenergic receptors are present in the carotid body and that such agonists exert an inhibitory influence on the chemoreceptor response to hypoxia [29]. Further, dexmedetomidine administered

intravenously to dogs resulted in a diminished response to increased CO₂, lasting for approximately 2 hours [30]. In agreement with earlier reports on α_2 -agonists [8,17], sedation with α_2 -agonists was associated with a significant increase in pulmonary and systemic arterial blood pressure. Although the distribution of blood flow from hypoxic regions in the lung to ventilated areas is highly efficient in the pony [22], it is possible that the elevated PAP may disturb this mechanism for matching of the perfusion to ventilated areas and thereby also contributes to impaired arterial oxygenation [31].

The slightly higher PaO₂ predicted by the multiple inert gas elimination technique (MIGET) compared to the measured PaO₂ may be due to diffusion limitation or extra-pulmonary reasons. Diffusion limitation can be caused by a limited gas equilibration time or by structural changes of the alveolar-capillary interface. Diffusion limitation seems unlikely as the cardiac output was not high enough to cause time limited gas equilibration and no clinical signs of pulmonary oedema were seen. Administration of the α_2 -agonist dexmedetomidine to dogs has been shown to decrease cardiac output with 50%, resulting in decreased perfusion of skin and muscle without decrease in blood flow to the heart [32]. Venous blood from the heart enters the arterial circulation through the Thebesian vein, without going through the lung and is not a part of the MIGET measurements. Thus, the difference between predicted and measured PaO₂ during detomidine sedation may be due to a proportionally larger contribution from the Thebesian vein to the arterial circulation which lowers the PaO₂.

A reduction in mixed venous PO₂ from 4.3 to 3.5 kPa accompanied the decrease in arterial oxygenation during detomidine sedation in the present study. A reduction in cardiac output decreases PvO₂ when oxygen consumption remains unchanged. Although there was a tendency for increased haemoglobin concentration and oxygen carrying capacity in the blood during detomidine sedation this effect was overridden by the pronounced decrease in cardiac output. The final result was an overall decrease in oxygen delivery to the tissue and increased oxygen extraction. The reduced PvO₂ further reduces PaO₂ for the same degree of ventilation-perfusion mismatch [33]. Thus, the slight but significantly increased V_A/Q mismatch measured during sedation in the present study further aggravated the pulmonary gas exchange, especially in the presence of impaired perfusion.

Detomidine and butorphanol combination

This drug combination is reported to have minimal effects upon the cardiovascular system [11] and usually does not cause any circulatory changes beyond those induced by the α_2 -agonist alone although there may be a

slight further respiratory depression [3,4]. In the present study, the only clear effect on pulmonary gas exchange by the combination of detomidine and butorphanol was a further decrease in ventilation, with additional increase in PaCO₂. This finding is probably an effect of butorphanol since the effect of the detomidine administered intravenously 35 minutes earlier is most likely diminished [17,28]. Lavoie et al. [5] found that a combination of detomidine and butorphanol in healthy horses as well as in horses with pre-existing respiratory dysfunction affected the respiratory function.

In the present study the increased P(A-a)O₂ persisted when butorphanol was additionally administered but the contribution of the causative factors changed. After butorphanol administration, the V_A/Q distribution improved and both Q_{mean} and V_{mean} were normalised. The shift of V_A/Q distribution to relatively lower but normal range was achieved by the reduction in ventilation, which now matched the reduced blood flow (Figure 1). Interestingly, the fraction of dead space ventilation was reduced compared to values during sedation with detomidine alone. This possibly reflects an improved distribution of blood flow, since vascular resistance was reduced compared to the values during detomidine sedation. This is in line with earlier investigation on sedation in the horse [17] that has showed a reduction in vascular resistance over time.

Conclusion

The results of the present study suggest that both pulmonary and cardiovascular factors contribute to the impaired pulmonary gas exchange during detomidine and butorphanol sedation in the horse. A significant reduction in blood flow and increase in V_A/Q maldistribution are the major contributors to the alveolar-arterial oxygen tension difference during sedation with detomidine. After addition of butorphanol P(A-a)O₂ remained impaired despite the improved V_A/Q distribution. This was caused by decreased ventilation, induced by the butorphanol administration, which matched a persistent low blood flow. No increase in intrapulmonary shunt compared to unsedated horses was evident during detomidine sedation or subsequent butorphanol administration.

Competing interests

SM is employed by Orion Pharma Animal Health, Sollentuna, Sweden. This investigation was carried out as a part of Marntell's PhD thesis at the Department of Clinical Sciences, Faculty of Veterinary Medicine and Animal Science, Swedish University of Agricultural Sciences, Uppsala, Sweden.

Authors' contributions

GN planned and organised the study and was in charge of the practical work. GN and SM collected and analysed data and prepared major parts of the manuscript. GH participated in interpretation of the pulmonary function and in critically revising the manuscript. AE, PF and KM contributed in collection of samples and the laboratory work as well as handling horses. All authors read and approved the final manuscript.

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References

- Virtanen R: **Pharmacology of detomidine and other alpha-2-adrenoceptor agonists in the brain.** *Acta Vet Scand Suppl* 1986, **82**:35–46.
- Ruffolo RRJ, Nichols AJ, Stadel JM and Hieble JP: **Pharmacologic and therapeutic applications of alpha 2-adrenoceptor subtypes.** *Annu Rev Pharmacol Toxicol* 1993, **33**:243–279.
- Clarke KW, England GCW and Goossens L: **Sedative and cardiovascular effects of romifidine, alone and in combination with butorphanol.** *J Assoc Vet Anaesth* 1991, **18**:25–29.
- Clarke KW and Paton BS: **Combined use of detomidine with opiates in the horse.** *Equine Vet J* 1988, **20**(5):331–334.
- Lavoie JP, Phan ST and Blais D: **Effects of a combination of detomidine and butorphanol on respiratory function in horses with or without chronic obstructive pulmonary disease.** *Am J Vet Res* 1996, **5**:705–709.
- Muir WW, Skarda RT and Sheehan W: **Hemodynamic and respiratory effects of a xylazine-acetylpromazine drug combination in horses.** *Am J Vet Res* 1979, **40**:1518–1522.
- Clarke KW and Hall LW: **"Xylazine"—a new sedative for horses and cattle.** *Vet Rec* 1969, **19**:512–517.
- Clarke KW and Taylor PM: **Detomidine: a new sedative for horses.** *Equine Vet J* 1986, **18**:366–370.
- England GCW, Clarke KW and Goossens L: **A comparison of the sedative effects of three 2-adrenoceptor agonists (romifidine, detomidine and xylazine) in the horse.** *J Vet Pharmacol Ther* 1992, **15**:194–201.
- Nilsfors L, Kvarn C, Kallings P and Carlsten J: **Cardiorespiratory and sedative effects of a combination of acepromazine, xylazine and methadone in the horse.** *Equine Vet J* 1988, **20**:364–367.
- Robertson JT and Muir WWWW: **A new analgesic drug combination in the horse.** *Am J Vet Res* 1983, **44**(9):1667–1669.
- Taylor PM, Browning AP and Harris CP: **Detomidine-butorphanol sedation in equine clinical practice.** *The Veterinary Record* 1988, **388**–390.
- Wagner PD, Saltzman HA and West JB: **Measurement of continuous distribution of ventilation-perfusion ratios: Theory.** *J Appl Physiol* 1974, **36**:588–599.
- Hedenstierna G, Nyman G, Kvarn C and Funkquist B: **Ventilation-perfusion relationship in the standing horse: An inert gas elimination study.** *Equine Vet J* 1987, **19**:514–519.
- Wagner PD, Naumann PF and Laravuso RB: **Simultaneous measurement of eight foreign gases in blood by gas chromatography.** *J Appl Physiol* 1974, **36**:600–605.
- Kirk RE: *Experimental design: Procedures for the behavioural sciences* Pacific Grove, CA, USA: Brooks/Cole Publishing Company; 31995.
- Wagner AE, Muir WW and Hinchcliff KW: **Cardiovascular effects of xylazine and detomidine in horses.** *Am J Vet Res* 1991, **52**:651–657.
- Nyman G and Hedenstierna G: **Ventilation-perfusion relationships in the anaesthetised horse.** *Equine Vet J* 1989, **21**:274–281.
- Rehder K, Knopp TJ, Sessler AD and Didier EP: **Ventilation-perfusion relationship in young healthy awake and anesthetized-paralyzed man.** *J Appl Physiol* 1979, **47**(4):745–753.
- Wagner PD, Laravuso RB, Uhl RR and West JB: **Continuous distribution of ventilation-perfusion ratios in normal subjects breathing air and 100% O₂.** *J Clin Invest* 1974, **54**:54–68.
- Dueck R, Rathbun M and Greenburg AG: **Lung volume and VA/Q distribution response to intravenous versus inhalation anesthesia in sheep.** *Anesthesiology* 1984, **1**:55–65.
- Elliott AR, Steffey EP, Jarvis KA and Marshal BE: **Unilateral hypoxic pulmonary vasoconstriction in the dog, pony and miniature swine.** *Respir Physiol* 1991, **85**:355–369.
- Hlastala MP, Bernard SL, Erickson HH, Fedde MR, Gaughan EM, McMurphy R, Emery MJ, Polissar N and Glenn RW: **Pulmonary blood flow distribution in standing horses is not dominated by gravity.** *J Appl Physiol* 1996, **81**:1051–1061.
- Reitemeyer H, Klein HJ and Deegen E: **The effect of sedatives on lung function in horses.** *Acta Vet Scand Suppl* 1986, **82**:111–120.
- Hamm D, Turchi P and Jochle W: **Sedative and analgesic effects of detomidine and romifidine in horses.** *Vet Rec* 1995, **13**:324–327.
- Latimer FG, Eades SC, Pettifer G, Tetens J, Hosgood G and Moore RM: **Cardiopulmonary, blood and peritoneal fluid alterations associated with abdominal insufflation of carbon dioxide in standing horses.** *Equine Vet J* 2003, **35**:283–290.
- Dantzker DR and D'Alonzo GE: **The effect of exercise on pulmonary gas exchange in patients with severe chronic obstructive pulmonary disease.** *Am Rev Respir Dis* 1986, **134**(6):1135–1139.
- Short EC, Matthews N, Harvey R and Tyner CL: **Cardiovascular and pulmonary function studies of a new sedative/analgesic (detomidine/domosedan) for use alone in horses or as a preanesthetic.** *Acta Vet Scand Suppl* 1986, **82**:139–155.
- Kou YR, Ernsberger P, Cragg PA, Cherniack NS and Prabhakar NR: **Role of alpha-2-adrenergic receptors in the carotid body response to isocapnic hypoxia.** *Respir Physiol* 1991, **83**:353–364.
- Sabbe MB, Penning JP, Ozaki GT and Yaksh TL: **Spinal and systemic action of the alpha 2 receptor agonist dexmedetomidine in dogs. Antinociception and carbon dioxide response.** *Anesthesiology* 1994, **80**(5):1057–1072.
- Marntell S, Nyman G, Funkquist P and Hedenstierna G: **Effects of acepromazine on pulmonary gas exchange and circulation during sedation and dissociative anaesthesia in horses.** *Veterinary Anaesthesia and Analgesia* 2005, **2**:83–93.
- Lawrence CJ, Prinzen FW and de Lange S: **The effect of dexmedetomidine on nutrient organ blood flow.** *Anesth Analg* 1996, **83**:1160–1165.
- West JB: *Pulmonary Pathophysiology -the essentials* Baltimore, MD, USA: Williams and Wilkins; 31987.

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