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SEMINAL CHANGES IN BOARS AFTER HEAT STRESS

By

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LARSSON, KJELL and STIG EINARSSON: *Seminal changes in boars after heat stress*. Acta vet. scand. 1984, 25, 57—66. — The object of the present study was to investigate the influence of elevated ambient temperature on sperm production, sperm morphology and composition of seminal plasma in boars. A total of 8 boars were used, 4 of them were exposed to 35°C, in a climate room, during 100 h and 4 served as controls and were kept at 20°C during 100 h in the climate room.

Ejaculate volume and total sperm count per ejaculate remained unaltered. An obvious decrease in sperm motility was seen in all heat exposed boars 15—21 days after the exposure. The most consistent increase in sperm abnormalities were proximal cytoplasmic droplets and abnormal sperm heads. The highest levels were found during the 4th week after exposure. All the sperm characteristics assessed had returned to normal levels at the end of the experimental period, which means 7—8 weeks after the end of exposure.

Only minor and inconsistent alterations were found in the seminal plasma components analysed and these changes were observed both in control and experimental boars.

boars; elevated ambient temperature; sperm morphology; semen biochemistry.

Exposure of boars to elevated ambient temperature has been reported to cause lowered fertility (*Wetteman et al.* 1977), decreased ejaculate volumes, total sperm counts and percentages of motile spermatozoa, and increased numbers of abnormal spermatozoa (*McNitt & First* 1970, *Wetteman et al.* 1977). *Einarsson* (1968) found lowered ejaculate volumes and reduced fertility of Swedish AI-boars that had been kept outdoors during the warm summer period. Similar results were reported of *Thibault et al.* (1966) and by *Signoret & du Mesnil du Buisson* (1968).

McNitt *et al.* (1972) found the normal testicular temperature of boars to be 35.7°C and the scrotum surface temperature was 33.1°C. Exposure of the boars to 40°C for 80 min caused a significant increase of the testicular temperature.

Local heating of the scrotum causes disturbances in spermiogenesis resulting in lowered sperm concentration, lowered percentages of motile spermatozoa and increased numbers of abnormal sperm cells (Holst 1949, Mazzari *et al.* 1968). The aforementioned studies indicate that exposure of boars to elevated ambient temperature causes similar morphological alterations in the ejaculates. The effects on accessory sex gland function have received little attention.

The aim of the present study was to investigate the influence of elevated ambient temperature on sperm production, sperm morphology and composition of seminal plasma in boars.

MATERIAL AND METHODS

Eight sexually mature Swedish Landrace boars were used in the study. The boars were brought to the clinic at approximately 7 months of age. At the clinic they were kept in individual pens, the ambient temperature being approximately 18°C. The boars were fed a conventional diet and none of them showed clinical signs of disease. Semen was collected twice weekly from all boars and all boars produced at least 8 ejaculates with normal sperm morphology, according to the standards of our laboratory, before exposure to elevated temperature.

Ten days before heat stress permanent vein catheters were inserted into one of the brachial veins according to the method of *Karlbom et al.* (1982). Blood samples were drawn at regular intervals through the cannulas during the whole experimental periods. The results from the hormone analyses have been published elsewhere (Larsson *et al.* 1983).

The boars were exposed to elevated temperature in a temperature controlled room (climate room) which was about 2.8 m square i.e. somewhat larger than the original pens. Duration of exposure was 100 h and the boars were then returned to their original pens. Four boars (experimental group) were exposed to 35°C and 40 % relative humidity and 4 boars (control group) were kept in the climate room at 20°C and 60—70 % relative humidity. Semen collections were not performed in the climate room.

Semen collections were performed by the gloved-hand technique and the boars were allowed to mount the dummy sow in their own pens. At collection the gel portion of the ejaculate was filtered away through double gauze and the remaining part of the ejaculate was collected into a plastic bag in a preheated vacuumflask. Ejaculate volumes were measured in graded glass cylinders, sperm motility was estimated in a phase-contrast microscope at 37°C and at a maximum of $\times 400$ magnification. Sperm morphology was assessed by phase-contrast microscopy of formol-saline fixed wet preparations (*Hancock 1957, Bane 1961*) and by light microscopy of Williams-stained dry smears (*Lagerlöf 1934*). Sperm concentration was measured by haemocytometric counting. Within 10 min after semen collection 20 ml of each ejaculate was centrifuged for 20 min, the seminal plasma was then analysed for fructose and the remaining part of the seminal plasma was stored at -20°C until analysed for concentrations of sodium, potassium, magnesium, chloride and total protein. The methods used for the biochemical analyses were the same as previously used by *Einarsson (1971)*.

All the boars were slaughtered at the end of the experimental period. The sexual organs were collected as soon as possible after slaughter. All sexual organs were carefully examined macroscopically. Specimen from each testicle were fixed in Bouin's fluid for histological examination. The pieces of tissue were dehydrated, embedded in paraffin, sectioned and stained with haemalum-eosin and van Gieson's haemalum picrotocin.

RESULTS

The results of the study are summarized in Tables 1—5. The values referred to as "before" represent mean and standard deviations for 8 ejaculates from each boar. The associated figures in the tables represent the number of boars, with a measured value different from the mean ± 2 s of the 8 ejaculates collected before exposure, on the particular occasion. Thus the comparisons between "before" and "after" exposure were made within boars and as combined figures for all boars.

Ejaculate volume, total sperm count per ejaculate and percentage of motile spermatozoa are presented in Table 1. No consistent differences in ejaculate volume and sperm count per

Table 1. Ejaculate volume, total sperm count per ejaculate and per cent motile spermatozoa in boars before and after treatment in climate room. The associated figures indicate numbers of boars with a measured value different from the mean ± 2 s of the ejaculates collected before exposure.

Days after exposure	Volume (ml)		Total sperm count ($\times 10^9$)		Motility (%)	
	Temp in climate room		Temp in climate room		Temp in climate room	
	35°C	20°C	35°C	20°C	35°C	20°C
Before	167 \pm 26	135 \pm 15	45.5 \pm 12.2	38.7 \pm 6.8	74 \pm 2	73 \pm 8
1—7	173	152	55.0	55.3 ²⁾	69 ¹⁾	71
8—14	204	172 ²⁾	56.9 ¹⁾	51.6 ¹⁾	68 ²⁾	73
15—21	179	163 ²⁾	55.5	49.4 ¹⁾	57 ⁴⁾	74
22—28	185	154 ¹⁾	51.7	55.1 ¹⁾	57 ²⁾	75
29—35	169	128	57.1 ¹⁾	63.1 ¹⁾	66 ²⁾	75
36—42	174		50.2		70 ¹⁾	
43—49	176		55.2 ²⁾		71	
50—56	196		69.8 ¹⁾		74	

ejaculate were observed within or between control boars and experimental boars, respectively, throughout the experimental period. The percentages of motile spermatozoa remained on pre-exposure levels in the control boars. In the ejaculates from the experimental boars an obvious decrease in sperm motility was seen in all 4 boars 15—21 days after the exposure. In 2 of these boars sperm motility was lowered already 8 days after exposure. The motility had returned to “before exposure” values in all 4 boars at day 43.

Tables 2 and 3 present the sperm morphology in the ejaculates before and after exposure. The most consistent increases of sperm abnormalities were proximal cytoplasmic droplets and abnormal sperm heads. Only minor or inconsistent increases of the other sperm abnormalities were observed in the experimental boars. No deviations in sperm morphology were seen in any of the control boars. The incidence of proximal cytoplasmic droplets increased continuously and was highest during the 4th week after exposure and thereafter decreased. The incidence of abnormal sperm heads started to increase markedly 15—21 days after exposure reaching its highest level at days 22—28. All the sperm characteristics assessed had returned to “before” levels at the end of the experimental period.

Table 2. Proximal cytoplasmic droplets, defective middle-pieces and bent or coiled tails in boars before and after treatment in climate room. The associated figures indicate number of boars with a measured value different from the mean ± 2 s of the ejaculates collected before exposure.

Days after exposure	Prox cytopl droplets (%)		Defective middle-pieces (%)		Bent or coiled tails (%)	
	Temp in climate room		Temp in climate room		Temp in climate room	
	35°C	20°C	35°C	20°C	35°C	20°C
Before	1.6 \pm 1.6	0.9 \pm 0.7	1.9 \pm 1.1	2.4 \pm 2.4	0.8 \pm 0.6	0.3 \pm 0.3
1—7	5.8 ¹⁾	1.0	1.3	1.8	0.8	0.6
8—14	13.3 ³⁾	0.7	1.6	1.4	1.1	0.2
15—21	22.4 ²⁾	0.8	2.6 ¹⁾	1.3	2.8	0.2
22—28	23.9 ³⁾	0.3	3.5 ²⁾	1.8	4.1	0.4
29—35	8.7 ²⁾	0.3	2.1 ¹⁾	0.9	1.8	1.6
36—42	4.9 ¹⁾		1.8 ¹⁾		0.8	
43—49	6.0 ²⁾		0.8		0.5	
50—56	3.5		0.8		0.5	

Table 3. Abnormal sperm heads, acrosome defects and tailless heads in boars before and after treatment in climate room. The associated figures indicate number of boars with a measured value different from the mean ± 2 s of the ejaculates collected before exposure.

Days after exposure	Abnorm sperm heads (%)		Acrosome defects (%)		Tailless heads (%)	
	Temp in climate room		Temp in climate room		Temp in climate room	
	35°C	20°C	35°C	20°C	35°C	20°C
Before	2.0 \pm 0.4	2.6 \pm 0.7	1.8 \pm 1.0	2.5 \pm 1.6	0.3 \pm 0.2	0.3 \pm 0.3
1—7	1.3	2.5	0.6	2.5	0.1	0.5
8—14	2.3	3.2	0.8	2.5	1.1	0.2
15—21	19.4 ³⁾	1.8	4.0 ²⁾	0.9	0.9 ²⁾	0.3
22—28	20.2 ⁴⁾	1.8	7.3 ¹⁾	0.9	1.2 ¹⁾	0.1
29—35	5.3 ³⁾	2.4	2.1	0.3	0.2	0.2
36—42	3.4 ¹⁾		1.5		0.2	
43—49	2.6		0.5		0.5	
50—56	2.0		0.5		0.3	

The results of the seminal plasma analyses are presented in Tables 4—5. Only minor and inconsistent alterations were found in the seminal plasma components and these changes were observed both in control boars and in experimental boars.

The morphological examination of the testicular tissues revealed no lesions in the experimental boars or in the control boars.

Table 4. Sodium, potassium and chloride in seminal plasma in boars before and after treatment in climate room. The associated figures indicate number of boars with a measured value different from the mean ± 2 s of the ejaculates collected before exposure.

Days after exposure	Sodium (mmol/l)		Potassium (mmol/l)		Chloride (mmol/l)	
	Temp in climate room		Temp in climate room		Temp in climate room	
	35°C	20°C	35°C	20°C	35°C	20°C
Before	130.8 \pm 3.1	130.8 \pm 6.0	13.9 \pm 0.4	14.0 \pm 0.9	104.9 \pm 5.3	106.2 \pm 8.7
1—7	131.1	130.1	14.4 ¹⁾	14.6 ¹⁾	104.8	103.8 ¹⁾
8—14	131.8	131.9	13.7	14.0	106.0	107.1
15—21	130.1	133.6	14.3 ¹⁾	14.0	106.5	108.1
22—28	129.6	140.8 ¹⁾	14.0 ²⁾	14.6 ¹⁾	102.9 ²⁾	117.0 ¹⁾
29—35	136.1 ¹⁾	131.3	15.2 ²⁾	14.7 ¹⁾	106.0 ¹⁾	104.7 ¹⁾
36—42	127.3		14.4		101.1	
43—49	127.0 ¹⁾		14.6		100.3 ¹⁾	
50—56	140.7 ¹⁾		16.1 ²⁾		115.8 ¹⁾	

Table 5. Fructose, magnesium and total protein in seminal plasma in boars before and after treatment in climate room. The associated figures indicate number of boars with a measured value different from the mean ± 2 s of the ejaculates collected before exposure.

Days after exposure	Fructose (mg/ejaculate)		Magnesium (mmol/l)		Total protein (g/l)	
	Temp in climate room		Temp in climate room		Temp in climate room	
	35°C	20°C	35°C	20°C	35°C	20°C
Before	31.4 \pm 18.3	41.2 \pm 41.6	4.93 \pm 0.23	4.10 \pm 1.77	16.9 \pm 2.4	15.1 \pm 3.9
1—7	33.6 ¹⁾	38.7	4.91 ¹⁾	4.26 ¹⁾	16.4	16.1
8—14	54.7 ¹⁾	40.2 ¹⁾	4.55	3.95	16.4	14.9
15—21	48.5 ¹⁾	39.3 ²⁾	5.01	3.73 ¹⁾	17.0	14.9
22—28	35.7 ¹⁾	15.1	5.21 ¹⁾	3.68	17.9 ¹⁾	15.6
29—35	36.8	16.5	5.98 ²⁾	2.58 ²⁾	21.6 ²⁾	12.8
36—42	23.3 ¹⁾		5.00		18.4 ¹⁾	
43—49	18.8 ¹⁾		4.97		18.4	
50—56	30.2 ¹⁾		4.48		17.3 ¹⁾	

DISCUSSION

During the post-exposure period neither total sperm counts nor ejaculate volumes of the boars were altered. *McNitt & First* (1970) found decreased sperm numbers in the ejaculates after heat exposure. The most evident changes were measured 28—32 days after exposure. The variations between ejaculates within

boars were great in the present study. It is therefore likely, that small variations due to treatment have been overlooked.

The lowered motility and the morphological alterations of the ejaculated spermatozoa for the boars that were exposed to elevated ambient temperature are in accordance with earlier studies (e.g. *Holst* 1949, *Mazzari et al.* 1968, *McNitt & First* 1970, *Einarsson & Larsson* 1982). However, since only minor and inconsistent alterations were found in spermatozoa from boars that were exposed for 24 h in the same climate room (*Einarsson & Larsson* 1982), it is likely that the duration of exposure is of importance for the disturbances. Alterations in the sperm morphology were found already 8 days after the end of the 100 h exposure, indicating that the epididymal influence on sperm maturation had been disturbed. The most consistent increase was found in the percentage of spermatozoa with proximal cytoplasmic droplets. According to *Holst* (1949) the first sign of disturbance in the spermiogenesis in the boar is an increased incidence of proximal cytoplasmic droplets. The migration of the cytoplasmic droplet normally takes place in a narrow region of the caput epididymis (*Gustafsson et al.* 1970) and the continuous increase of spermatozoa with proximal cytoplasmic droplets might therefore indicate a disturbed epididymal function. However, the percentages of spermatozoa with single-bent or coiled tails, an indicator of epididymal function (*Einarsson & Gustafsson* 1973, *Bonte et al.* 1978) were low throughout the experimental period. Thus, the exposure to elevated temperature did not induce a genuine epididymal dysfunction. The cause of the morphological changes of the spermatozoa, which appeared very early after heat exposure, might be that epididymal function is partly affected by scrotum temperature and/or by the hormonal changes that occur during and after exposure to elevated ambient temperature (*Larsson et al.* 1983).

In the boars that were exposed to elevated ambient temperature the incidence of abnormal sperm heads increased during the postexposure period, indicating a disturbed spermiogenesis. It might be postulated that all spermatozoa collected ≤ 7 days after the end of exposure to elevated ambient temperature had left the testicles already at the beginning of exposure, while very few of the spermatozoa collected 15—21 days after exposure had left the testicles during exposure (*Swierstra* 1968). Only in 2 of the 4 boars were very high percentages of abnormal sperm heads

found. In these 2 cases were also the incidences of proximal cytoplasmic droplets very high. No indication of a decreased sperm production was found in any boar in the present investigation. The testicular disturbance must therefore be considered as slight or moderate but in no case severe. All the sperm characteristics measured had returned to normal levels at the end of the experimental period. Furthermore no lesions were observed in the testicular tissues collected at the end of the experimental period, which indicate that a complete regeneration already had taken place.

The results of the analyses of the concentrations of the seminal plasma components before and after heat exposure did not indicate any changes. The non-responses of the levels of total protein and fructose to the treatment might indicate that neither the decreased testosterone levels during the heat exposure nor the elevated testosterone levels during the first 5 days after heat exposure did influence the activity of the accessory sex glands. Identical results were presented by *Egbunike & Jeyakumar* (1980). In their study boars, adapted to natural tropical conditions, were subjected to direct heat stress by exposure to direct sunlight. The seminal plasma protein content and the fructose levels remained unchanged. The protein profile was however different between heat-stressed boars and control boars. As the quality of the proteins in the seminal plasma was not evaluated in the present study it is impossible to certify that the accessory glands were unaffected by the heat exposure.

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SAMMANFATTNING

Spermaförändringar av värmestress hos galtar.

Ändamålet med denna undersökning var att studera inverkan av förhöjd omgivningstemperatur på spermaproduktion, spermiemorfologi och spermaplasmans sammansättning hos galtar. Totalt ingick 8 galtar i försöket. Fyra av dem exponerades för 35°C under 100 timmar i en klimatkammare, medan 4 galtar användes som kontroller med 20°C temperatur under 100 timmar i klimatkammare.

Ejakulatvolym och totalantal spermier per ejakulat påverkades inte. Spermimotoiliteten sjönk hos alla galtar som utsattes för värmestress och var lägst 15—21 dagar efter värmestressen. De mest påtagliga förändringarna i spermiemorfologin var förhöjda frekvenser proximala cytoplasmadroppar och spermiehuvuddefekter. De högsta nivåerna återfanns under 4:e veckan efter värmestressen. Spermimorfologin hade normaliserats vid slutet av försöksperioden d v s 7—8 vecor efter värmestressen.

Spermaplasmans sammansättning var i stort sett oförändrad under hela försöksperioden. Mindre förändringar observerades hos såväl kontrollgaltar som försöksgaltar.

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