Bacteriological and Histological Investigation of the Postpartum Bovine Uterus in Two Estonian Dairy Herds

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> Kask K, Kindahl H, Gustafsson H: Bacteriological and histological investigation of the postpartum bovine uterus in two estonian dairy herds. Acta vet. scand. 1998, 39, 423-432. - Postpartum uterine infections, endometrial histology and resumption of ovarian activity in cows were studied in 2 Estonian dairy herds with different herd sizes, milk yields and management systems. Ten cows at Farm A and 5 cows at Farm B were studied in the experiment. All cows in the study had normal calving performance. Endometrial biopsies for bacteriological and histological examinations were collected once a week starting on the second week postpartum and continuing for 7 weeks postpartum. Milk progesterone samples were collected twice a week during the whole study period. In both herds, the uterine flora contained mainly facultative anaerobic bacteria (Streptococcus spp., E. coli, Staphylococcus spp., Proteus vulgaris). Among obligate anaerobic bacteria only Bacteroides spp. were found. After 7 weeks of collection at farm A, a bacterial uterine flora still persisted in 2 of the cows. At farm B, on the other hand, bacterial elimination was complete after 6 weeks. Presence of inflammatory cells in uterine histology specimens remained higher at the end of collection and resumption of ovarian activity was delayed at farm A. After 7 weeks postpartum, only 6 of the 10 cows at farm A had resumed ovarian cyclicity, while at farm B the first oestrous cycle had occurred in all cows. The study showed that differences regarding uterine infections and their clearance occured between farms and, despite these differences, cows with normal calving performance will effectively recover without any treatment.

cow; uterine bacteriology; uterine histology; ovarian activity.

Introduction

Cattle breeding in Estonia has always been based on milk production. Milk and milk products form an important part of human nutrition and are the main agricultural export products at the moment. Dairy cattle production makes up about 60% of the national agricultural commodity production. The total number of cows was 173 200 in December 1996, and the average milk production was 3 913 kg. The potential annual production in Estonia is, however, 6000-8000 kg milk per cow. These production levels have already been achieved in some herds, where feeding and management are outstanding.

According to official animal records (1996), the reasons for culling cows in Estonian herds are foremost fertility problems (33.9%). Many of the fertility problems are linked to suboptimal

management factors and feeding of the cows. Many farmers and veterinarians believe that the use of preventive antibiotic treatments can compensate for poor management. This is also the case where there is no obvious need for the treatment. There are some farms where all animals were treated in the postpartum period using antibiotics and hormonal drugs. In the future, such treatments should be avoided for animal health reasons and in order to decrease pharmacological residues in meat and milk.

The postpartum period (pp) is important in the life of the dairy cow. During this period the animal should eliminate intrauterine infections, that may be present, and re-establish normal uterine and ovarian functions. Diseases such as endometritis, mastitis and ketosis are common during pp and some of the problems are caused by micro-organisms. The intrauterine micro-organisms isolated during pp from the uterus have been described in many early studies and have been found to prolong the uterine involution and also very often lead to early culling of the cows (Roberts 1967, Bostedt 1984, Fredriksson et al. 1985, Kındahl et al. 1986, Bekana 1996). The risk for uterine infections increases in cows with abnormal calving or retained fetal membranes (RFM) (Ruder et al. 1981a, Olson et al. 1984, Fredriksson et al. 1985, Markusfeld 1993, Bekana et al. 1994a,b).

The present study was conducted on 2 Estonian dairy farms of different size and milk yield and with different management systems. The aims were to study the postpartum period of cows with normal calving. The main emphasis was placed on uterine infections, but also the re-establishment of ovarian cyclicity was studied, using milk progesterone analyses. An important goal was also, based on the results of the study, to give information to Estonian veterinarians and farm owners about treatment regimes for postpartum cows.

Materials and methods

Farms

Two farms (A and B) both belonging to the State farm in Tartu County, were studied. Some data on the farms are given in Table 1. Feeding standards on both farms were according to the milk yield of the cows. The rations in farm A consisted of hay, straw, silage and fodder beet. In farm B the study was conducted in late summer and rations were based on pasture and other green feeds. On both farms, some concentrates were also given, the origin of which was not industrial and consisted mainly of mixtures of different grains made on the same state farm. On both farms the cows were given 400 g of concentrate per liter of daily milk yield.

Animals

Fifteen Estonian dairy cows were used in the experiment, 10 of which belonged to farm A and 5 to farm B. They were of Estonian black and white breed (EBW) and of Estonian red breed (ER). Animals on farm A calved on March 12, 1996 (n = 5) and March 13, 1996 (n = 5). None had a difficult calving. One animal had retention of placenta for 24 h after calving. Animals from farm B calved on September 1, 1996 (n = 3) and September 3, 1996 (n = 2), none having difficult calvings or retained fetal membranes. No treatment was given to the animals either before or after calving. This group of animals was at pasture during daytime.

Collection of specimens for bacteriology and histology

Each animal in this study was sampled for bacteriological and histological examination once a week, starting within 10 days after parturition and continuing for 7 weeks on farm A and for 6 weeks on farm B. Endometrial biopsies were aseptically collected according to the techniques and methods described by *Fredriksson et al.* (1985) and *Bekana et al.* (1994b).

Farm	No cows	Breeds	Mılk prod kg	Mılkıng	Housing, management				
Α	601 ER 4900 SLB EBW	4900	2× per day, machine pipeline	Tying system. Removal of manure by tractor scraper once a day. Feeding also by tractor. Routine treatment of endometritis using antibiotic					
В	394	EBW SRB	5200	2× per day, machine pipeline	Tying system. Removal of manure twice a day by an electric scraper. Feeding manually from wagons. Routine use of intrauterine antibiotic treatment in most of the postpartum cows.				

 Table 1
 Characteristics of 2 Estonian farms used in the study

ER = Estonian red breed; EBW = Estonian black and white breed, SLB = Holstein Friesian breed; SRB = Swedish red and white breed

The cow was restrained, the faeces removed from the rectum and the tail was secured, the perineal area and vulva were washed carefully and the vulvar area and the vestibulum were disinfected using a mild iodophore. Then the vulvar lips were parted to introduce an outer protective stainless steel tube. The instrument was fixed in the external opening of the cervix. A long guarded culture instrument was advanced into the uterus by cervical manipulation (Messier et al. 1984, Fredriksson et al 1985). The instrument was opened and a milled cavity on the tip of the instrument was located with the forefinger of the hand in the rectum. This cavity, with its sharpened edge, formed a curette by which biopsies were obtained from the endometrium.

Biopsies for bacteriological examination were immediately placed in thioglycolate medium for transportation to the laboratory for bacteriological examination. Samples were cultured within 2 h. Isolation of the bacterial species was performed at the Department of Veterinary Microbiology, Estonian Agricultural University, Tartu using standard bacteriological procedures. Plates cultivated aerobically were examined after 24 h and 48 h and plates cultivated anaerobically after 48h and 168 h. All bacterial isolates were identified according to Bergey's Manual of Systematic Bacteriology (*Holt* 1986). The results were intepreted as follows:

- 0 = no bacteria;
 - 1 = mild growth;
 - 2 =moderate growth;
 - 3 = heavy growth.

Biopsies for histological examination were placed in 10% formol salue solution. After fixation, tissues were trimmed, embedded in paraffin, sectioned at 6µm and stained with haematoxylin and eosin. All the procedures were made in the Histology laboratory, Department of Obstetrics and Gynaecology, Swedish University of Agricultural Sciences in Uppsala. Investigations were made under the microscope for presence of inflammatory cells in the endometrium. Infiltration of the neutrophils in the biopsy specimen was considered as normal to slight, medium or dense. The diffuse cellular reaction involving also the mononuclear cells was assessed by counting the inflammatory cells in 6 high power fields chosen randomly and calculating the average number of cells per high power field (Griffin et al. 1974). The results were interpreted as follows:

Normal to slight infiltration = 0 to 29 cells per field;

Medium infiltration = 30 to 80 cells per field; Dense infiltration = more than 80 cells per field.

Progesterone

Milk samples for progesterone determination were collected twice weekly, starting on the same day as the first biopsies were taken and continuing until the end of the study. Milk sampling was performed within 60 min after the morning milking. Samples from each cow were collected into plastic tubes containing $100 \,\mu$ l of preservative (Bronopol 2% + MTB 0.05%) and stored at 4 °C until assay. Analyses were performed at the Department of Clinical Chemistry, Swedish University of Agricultural Sciences in Uppsala. The content of milk progesterone was determined by RIA (Spectria, Orion Diagnostica, Espoo, Finland). The interassay coefficient was below 14% and the intraassay coefficient was below 17%. Detection limit of the assay was 1 nmol/l. The minimum progesterone value for detection of luteal activity was considered to be 6.7 nmol/l (Laitinen 1983).

Statistical analyses

The statistical analyses were made using the Statistical Analysis System (*SAS Institute Inc.* 1994). The FREQ procedure for Chi-Square test and Mantel-Haenszel Chi-Square test were used. p<0.05 was considered as a significant difference.

Results

The overall results from bacteriological and histological examinations are presented in Table 2.

Bacteriology

The results of bacteriological examination are presented in Tables 2 and 3. Altogether 94 biopsies were collected from the 15 animals. From these, 45 were found to be positive and the remaining 49 biopsies negative. Six samples are missing due to problems during collection. Five samples from the positive biopsies showed mixed infections and the remaining 40 samples were facultative anaerobic bacteria in pure culture. The mixed cultures contained mainly *Bacteroides* spp., *Escherichia coli* and *Streptococcus* spp. In one case also *Proteus vulgaris* was involved. A total of 58 isolates were identified among the 45 bacteriologically positive biopsies. From these, 5 were obligate anaerobic bacteria. The most common anaerobic pathogens were *Streptococcus* spp. *E. coli* and *Staphylococcus* spp. The only obligate anaerobic bacteria found were *Bacteroides* spp. (Table 3).

The highest incidence of bacteriological growth, and also the highest number of bacterial species, was found during the first 4 weeks after parturition. Elimination of the bacterial isolates is shown in Figs. 1 and 2 (Farm A versus B). After eight weeks postpartum, 1 out of 15 animals still remained bacteriologically positive (Table 2). No totally negative animals were found during the collection time at either farms, but the largest number of bacteriologically positive samples was seen on farm A. No significant difference between farms was detected (p>0.05).

Histology

Altogether 92 biopsies were collected for histology. A total of 8 biopsies are missing due to difficulties in collection. The results of the histological examinations, according to the farms, are presented in Table 2, and Figs. 1 and 2. Arithmetic means of the real numbers of the inflammatory cells and the percentage of cows with cell infiltration are given simultaneously in the figures. No significant difference between the 2 farms was detected (p>0.05).

Progesterone

On farm A, 6 out of 10 animals showed resumption of ovarian cycles during the first 8

	Bacteriology during 2-8 weeks pp							Histology during 2-8 weeks pp						
Cow No	2	3	4	5	6	7	8	2	3	4	5	6	7	8
Farm A:														
2486	3	2	0	0	0	0	?	D	D	Μ	?	Μ	Μ	Μ
9933	3	2	2	0	0	0	?	?	Μ	Μ	Μ	S	S	S
9479	3	3	3	3	2	2	1	D	Μ	?	Μ	Μ	Μ	Μ
2382	2	3	2	0	0	0	?	Μ	?	D	Μ	Μ	Μ	S
2703	3	0	1	0	0	0	?	Μ	Μ	Μ	S	S	S	S
2972	3	3	3	3	0	0	?	Μ	S	D	Μ	Μ	S	?
2881	0	2	2	2	3	0	0	D	Μ	Μ	Μ	Μ	Μ	Μ
9606	3	1	2	2	0	0	0	Μ	Μ	Μ	Μ	D	Μ	D
2988	3	3	2	0	0	0	0	Μ	Μ	Μ	Μ	?	D	S
2471	3	2	0	0	0	0	?	Μ	S	Μ	S	S	?	?
Farm B:														
2348	0	3	2	1	0	0		D	Μ	Μ	Μ	Μ	S	
4283	0	2	2	0	0	0		Μ	Μ	Μ	S	S	S	
1449	3	0	0	0	0	0		D	Μ	Μ	Μ	Μ	S	
1311	3	3	2	1	0	0		Μ	Μ	Μ	Μ	Μ	S	
1464	2	0	0	0	0	0		М	Μ	Μ	S	S	S	

Table 2. Bacteriological and histological results during the study period in individual animals on farm A and farm B.

0 = no bacteria; 1 = mild growth; 2 = moderate growth; 3 = heavy growth; ? = missing sample; S = normal to slight infiltration; M = medium infiltration; D = dense infiltration.

	Week Postpartum									
Bacteria	2	3	4	5	6	7	8			
Farm A $(n = 10)$										
Facultative anaerobic:										
Streptococcus spp.	3	2	2	3	1	1	0			
E. coli	1	3	2	0	1	0	0			
Staphylococcus spp.	2	3	3	1	0	0	1			
Proteus vulgaris	4	0	0	0	0	0	0			
Pseudomonas aeruginosa	1	1	1	0	1	0	0			
Others	2	2	1	1	0	0	0			
Total = 43	13	11	9	5	3	1	1			
Obligate anaerobic:										
Bacteroides spp.	2	2	1	0	0	0	0			
Total = 5	2	2	1	0	0	0	0			
Farm $B(n=5)$										
Facultative anaerobic:										
Streptococcus spp	2	1	1	1	0	0				
E coli	0	2	2	0	0	0				
Staphylococcus spp.	1	0	0	0	0	0				
Total = 10	3	3	3	1	0	Õ				

Table 3. Presence and elimination of bacteria (number of isolates) from the uterus in 15 cows on 2 different farms starting on the second week postpartum and continuing 7 and 6 weeks postpartum.



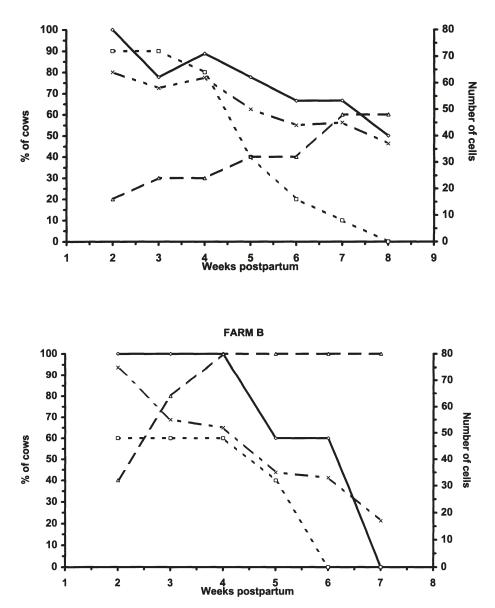


Figure 1 (farm A) and Figure 2 (farm B). Results of uterine bacteriological (uterine infections) (- - - \Box) studies and resumption of ovarian activity (— Δ) in 10 cows on farm A and 5 cows on farm B starting in the second week postpartum and continuing 7 weeks postpartum (6 weeks on farm B) Arithmetric means of cell numbers (— — X), (right Y-axis), and percentage of cows with medium or dense cell infiltration in the endometrium (—— Δ) are given simultaneously.

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weeks postpartum (values more than 6. nmol- l^{-1}). In 4 animals progesterone levels were found to be low during the whole period. On farm B, all 5 animals showed resumption of ovarian cyclicity during the 7 weeks period. Three of them started cyclicity in the third week postpartum. Results are presented in Figs. 1 and 2.

Discussion

The early studies from the sixties and seventies indicate that bacteria have been recovered from up to 93% of all cows during the early postpartum period (Elliott et al. 1968, Griffin et al. 1974). Later studies (Messier et al. 1984, Fredriksson et al. 1985, Bekana et al. 1996) have shown that intrauterine infections are not commonly found in cows with normal calving. In the present study, the methodology of Fredriksson et al. (1985) and Bekana et al. (1994b) was used. The results of bacteriological examinations of endometrial tissue biopsies are similar in principal when comparing these later studies, but they differ between bacterial species found. In the present study, the uterine flora contained mainly facultative anaerobic bacteria, but in no case was found Actinomyces pyogenes. This bacteria is the most commonly isolated bacteria in almost all postpartum bacteriological studies and has been considered as one of the most pathogenic agents, together with Fusobacterium necrophorum and Bacteroides spp. (Hartigan et al. 1974, Hartigan 1978, Ruder et al. 1981b, Fredriksson et al. 1985, Bekana et al. 1994b, Kaneko et al. 1997). Also, in a similar experiment conducted in Sweden at the same time and using also healthy animals, A. pyogenes was found to be one of the dominating isolates during the first 3 weeks postpartum, together with Bacteroides spp and F. necrophorum (Kask et al. 1998).

Among obligate anaerobic bacteria in the pre-

sent study, only Bacteroides spp. were found in 5 cases. In contrast to the Swedish study (Kask et al. 1998), the present study showed no totally negative animals. There was a tendency on farm B for fewer bacteria to be found during the collection period compared with farm A. The difference might be due to management factors such as unhygienic conditions before and after parturition when negative pressure is induced in the birth canal, and the birth canal is open for a period (Arthur et al. 1996). There were different hygiene conditions between the farms, with a better situation on farm B, where manure was removed twice a day. On farm A, removal was done usually only once a day. Unhygienic conditions in and around the cow may increase the bacterial contamination of the vestibulum and vagina, from where bacteria can easily migrate to the uterus after parturition. Especially E. coli, which was one of the dominating bacteria in this study, will mostly enter the uterus in such a way (Bretzlaff et al. 1982).

Generally, most of the non-specific bacteria found in this study, *Streptococci, Staphylococci* and *E. coli*, are short-lived, since the natural defence mechanism of the cow normally copes with invading bacteria. Thus the bacteria make sporadic and transient appearances in the uterine lumen without affecting subsequent fertility (*Griffin et al.* 1974, *Hartigan* 1978, *Ruder et al.* 1981a).

The highest incidence of isolates was detected during the first 5 weeks postpartum, after which the number decreased and at the end of the experiment (8 weeks postpartum) only one animal remained positive. This animal belonged to farm A and also had delayed delivery of placenta (24 h). This might have been a cause of delayed elimination of bacteria in that particular animal.

Histological results differed between farms and there were also differences between the time for bacterial elimination and normal histology. Af430

ter 8 weeks postpartum on farm A, medium and dense inflammatory cell infiltration was still found. The situation was better on farm B, where all 5 animals only had normal to slight infiltration of inflammatory cells in the endometrium by week 7 postpartum. According to *Skjerven* (1956), slight infiltration cannot affect subsequent fertility. On farm A, samples were missing during the last week in 3 animals, but 6 out of 10 animals had different degrees of infiltration. One animal still had a dense infiltration but without bacteria after 5 weeks. Also many animals that were still bacteriologically positive had only slight infiltration in the endometrium.

According to the progesterone values, 6 out of 10 animals on farm A showed resumption of ovarian cycles during the first 8 weeks postpartum. On farm B, all animals had started cycling by week 7 postpartum. There is a link between uterine involution and resumption of ovarian activity (Lamming et al. 1982, Kindahl et al. 1984, Butler et al. 1989). Of the animals with low levels of progesterone during the whole experimental period, one animal had dense, one animal medium and 2 animals had normal to slight infiltration in the endometrium at the end of experiment. On the other hand, among those animals that started cyclicity there were also cases of medium infiltration, especially on farm B. The reason might be a negative energy balance of the animals on Farm A. It is well known that negative energy balance after calving can cause delayed resumption of ovarian activity (Karg et al. 1982, Hanzen 1986, Imakawa et al. 1987). There are also suggestions that reduced resistance, which is related to negative energy balance, may play a role in delayed return of normal uterine function (Elliott et al. 1968). However, nutritional studies were not included within the scope of this study.

Conclusions

There were different hygienic conditions on the 2 farms in the experiment, and this seems to influence the results of the present study. Cows from farm B showed a tendency for less bacteriological contents and less histological changes in the uterus, and earlier resumption of ovarian activity, as compared with animals on farm A. Another important conclusion is that, despite the different hygienic conditions, all cows were recovering without any treatment. This leads to the suggestion that cows with normal calving performance do not need any antibiotic treatment.

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References

- Arthur GH, Noakes DE, Pearson H, Parkinson TJ Veterinary Reproduction and Obstetrics W B Saunders Company Ltd 1996, pp. 389-396.
- Bekana M Clinical, ultrasonographic, bacteriological and hormonal studies in post-partum cows with particular emphasis on retained fetal membranes Uppsala, Sweden 1996. PhD Thesis.
- Bekana M, Ekman T, Kındahl H Ultrasonography of the postpartum uterus with retained fetal membranes. J. Vet. Med. A. 1994a, 41, 653-662.
- Bekana M, Jonsson P, Ekman T, Kındahl H Intrauterine bacterial findings in postpartum cows with retained fetal membranes J Vet. Med A. 1994b, 41, 663-670
- Bekana M, Jonsson P, Kındahl H Intrauterine bacterıal findings and hormonal profiles in post-partum cows with normal puerperium. Acta vet. scand 1996, 37, 251-263.
- Bostedt H Uterine infections in the postpartum period. 10th International Congress on Animal Reproduction and AI, Urbana Champaign Illinois, June 10-14, 1984, Vol IV, III-25-III-33.

- Bretzlaff KN, Whitmore HL, Spahr SL, Ott RS. Incidence and treatments of postpartum reproductive problems in a dairy herd. Theriogenology 1982, 17, 527-535.
- Butler WR, Smith RD. Interrelationships between energy balance and postpartum reproductive function in dairy cattle. J. Dairy Science 1989, 72, 767-783
- *Elliott L, McMahon KJ, Gier HT, Marion GB*. Uterus of the cow after parturition: bacterial content. Am. J vet. Res. 1968, *29*, 77-81
- Fredriksson G, Kindahl H, Sandstedt K, Edqvist L-E: Intrauterine bacterial findings and release of $PGF_{2\alpha}$ in the postpartum dairy cow. Zbl Vet. Med. A. 1985, 32, 368-380.
- Griffin JFT, Hartigan PJ, Nunn WR: Non-specific uterine infection and bovine fertility. 1. Infections patterns and endometritis during the first seven weeks postpartum. Theriogenology 1974, 1, 91-106.
- Hanzen CH. Endocrine regulation of postpartum ovarian activity in cattle: a review. Reprod. Nutr. Dev. 1986, 26, 1219-1230.
- Hartigan PJ The role of non-specific uterine infection in the infertility of clinically normal repeatbreeder cows. Vet. Science Comm. 1978, 1, 307-321.
- Hartigan PJ, Griffin JFT, Nunn WR Some observations on Corynebacterium pyogenes infection of the bovine uterus. Theriogenology 1974, 1, 153-167
- *Holt JG* Bergey's Manual of Systematic Bacteriology Vol. 2, Williams and Wilkins Co, Baltimore, 1986.
- Imakawa K, Day L, Zalesky DD, Clutter A, Kittok RJ, Kinder JE Effects of 17beta-estradiol and diets varying in energy on secretion of luteinizing hormone in beef heifers. J. Anim. Sci. 1987, 64, 805-815.
- Kaneko K, Kawakami S, Miyoshi M, Abukawa T, Yamanaka S, Mochizuki M, Yoshihara S Effect of retained placenta on subsequent bacteriological and cytological intrauterine environment and reproduction in Holstein dairy cows. Theriogenology 1997, 48, 617-624.
- Karg H, Scallenberger E. Factors influencing fertility in the postpartum cow. Current Topics Vet. Med. Anim. Sci. Vol. 20 Martinus Nijhoff Publ., Boston 1982, MA. 63-72.
- Kask K, Gustafsson H, Magnusson U, Bertilsson J, Kindahl H Uterine bacteriology, histology and granulocyte function of the postpartum cows in

different milking frequencies. Submitted for publication 1998.

- Kindahl H, Cort N, Fredriksson G, Edqvist L-E, Stabenfeldt G Some aspects of the possible roles of diseases in altering reproductive performance. Proceedings of an International Symposium on the Use of Nuclear Techniques in Studies of Animal Production and Health in Different Environments jointly organized by the International Atomic Energy Agency and FAO of the United Nations, Vienna, 17-21 March, 1986, pp. 263-273.
- Kindahl H, Fredriksson G, Madej A, Edqvist L-E Role of prostaglandins in uterine involution. 10th International Congress on Animal Reproduction and AI, Urbana-Champaign Illinois, June 10-14, 1984, Vol IV, CI-9-XI-16
- Laitmen J Oestrus confirmation, pregnancy diagnosis and postpartum ovarian follow-up of the Finnish dairy cows by milk progesterone assay. Thesis 1983, University of Kuopio, Finland.
- Lamming GE, Peters AR, Rıley GM, Fisher MW Endocrine regulation of postpartum functions. Current Topics Vet Med Anim. Sci. 1982, 20, Martinus Nijhoff Publ., Boston, MA. 148-160.
- Markusfeld O. Parturition disease complex of the high yielding dairy cow Acta vet. scand. Suppl. 1993, 89, 9-15.
- Messier S, Higgins R, Couture Y, Morin M Comparison of swabbing and biopsy for studying the flora of the bovine uterus. Can. Vet. J. 1984, 25, 283-288.
- Olson J, Ball L, Mortimer RG, Farin PW, Adeny WS, Huffman M. Aspects of bacteriology and endocrinology of cows with pyometra and retained foetal membranes Am. J. vet. Res. 1984, 45, 2251-2255.
- Roberts DS The pathogenic synergy of Fusiformis necrophorus and Corynebacterium pyogenes II. The response of *F necrophorus* to a filterable product of *C pyogenes* Br. J. Exp. Pathol. 1967, 48, 674-679.
- Ruder CA, Sasser RG, Williams RJ, Ely JK, Bull RC, Butler JE. Uterine infection in the postpartum cow. effect of dietary crude protein restriction. Theriogenology 1981a, 15, 561-572.
- Ruder CA, Sasser RG, Williams RJ, Ely JK, Bull RC, Butler JE Uterine infections in the postpartum cow: possible synergistic effect of Fusobacterium necrophorum and Corynebacterium pyogenes Theriogenology 1981b, 15, 573-579.
- Skjerven O Endometrial biopsy studies in reproduc-

tively normal cattle (Clinical, histochemical and histological observations during the oestrus cycle). Acta endocrinologica 1956, Supplementum 26 (PhD thesis)

Sammanfattning

Bakteriologisk och histologisk undersökning av livmodern hos nykalvade kor i två estländska mjölkkor besattningar

Livmoderinfektioner, endometriehistologi och äggstocksaktivitet studerades hos mjölkkor i 2 estländska besättningar av olika storlek och med olika produktions- och skotselnivåer. Tio kor (besättning A) och 5 kor (besättning B) studerades i försöket. Alla kor i studien hade en normal kalvning och var inte föremål för någon behandling. Endometriebiopsier för bakteriell och histologisk undersökning togs ut en gång per vecka med borjan andra veckan efter kalv-

ning och 7 veckor framåt. Mjolkprov för progesteronanalys togs 2 gånger per vecka under försoksperioden. I båda besättningarna bestod bakteriefloran främst av fakultativt anaeroba bakterier (Streptococcus spp, E coli, Staphylococcus spp, Proteus vulgaris) Bland obligat anaerobik bakterier återfanns bara Bacteroides spp. I besättning A återfanns fortfarande bakterier i livmodern hos 10% av korna efter 7 veckors provtagning medan korna i besattning B var fria från bakterier efter 6 veckor Hos korna i besattning A noterades inflammatoriska celler i endometriet i hogre grad och senare igångsättning av äggstocksaktiviteten jamfört med korna i besättning B. Bara 60% av korna i besattning A hade haft sin första ovulation 7 veckor efter kalvning medan alla kor ovulerat i besattning B Studien visar att det föreligger skillnader mellan besättningar avseende elimination av livmoderinfektioner efter kalvning men att kor med normal kalvning, blir fria från bakterieinfektioner utan någon behandling

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