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# A Study on the Effects of Inhibition of Prostaglandin Biosynthesis with Flunixin Meglumine and Later Administration of Prostaglandin $F_{2\alpha}$ on the Intraluminal Pressure Variations in the Isthmus of the Oviduct in Unrestrained Gilts

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Pettersson, A., S. Einarsson and H. Kindahl: A study of the effects of inhibition of prostaglandin biosynthesis with flunixin meglumine and later administration of prostaglandin  $F_{2\alpha}$  on the intraluminal pressure variations in the isthmus of the oviduct in unrestrained gilts. Acta vet. scand. 1993, 34, 125-131. – Three gilts were each equipped with 2 ultra-miniature pressure sensors, placed at 2 different points along the same isthmus of the oviduct. Following base recordings of isthmic intraluminal pressure, the gilts were treated with 2.2 mg flunixin meglumine (FM) per kg body weight. After FM treatment, the peripheral plasma levels of 15-ketodihydro-PGF<sub>2a</sub>, the major metabolite of prostaglandin  $F_{2\alpha}$  (PGF<sub>2a</sub>), decreased within 30 min. The frequency of the phasic pressure fluctuations in the isthmus of the oviduct decreased after FM treatment. Exogenous administration of PGF<sub>2a</sub> increased the peripheral plasma levels of 15-ketodihydro-PGF<sub>2a</sub> produced an increase in the frequency of the phasic pressure fluctuations in the oviductal isthmus. When the PGF<sub>2a</sub> dose was increased to 0.5 mg, a marked increase in the base and total pressures was seen in addition to the increase in the frequency of the phasic pressure fluctuations.

porcine; oviductal isthmus.

# Introduction

The recording of intraluminal pressure variations can be used as an indirect method for studying muscle contractions in tubular organs.

Cyclic variations of the intraluminal pressure in the porcine oviductal isthmus have been demonstrated in vitro (*Rodriguez-Martinez et al.* 1982 a) and in vivo both in anaesthetized (*Rodriguez-Martinez et al.* 1982 b) and unrestrained gilts (*Pettersson* 1991). In unrestrained gilts, intraluminal pressure tended to increase through procestrus and the first day of standing cestrus (day 1), after which it tended to decrease, with low pressures recorded on days 3 and 4 (*Pettersson* 1991). The regulatory mechanisms of these cyclic variations in intraluminal pressure are still not fully understood. Exogenous administered prostaglandin  $F_{2\alpha}$  (PGF<sub>2\alpha</sub>) has a contractile effect on the porcine myosalpinx, both in vitro, and in anaesthetized gilts, in vivo (*Rodriguez-Martinez & Einarsson* 1985). The purpose of this study was to see if the intraluminal pressure in the isthmus of the oviduct in unrestrained gilts could be affected by a decreased endogenous prostaglandin production brought about by treating the gilts with the non-steroidal antiinflammatory drug flunixin meglumine (FM) and if administration of  $PGF_{2\alpha}$  could reverse any such effect.

### Materials and methods

# Animals

Three cycling mature Swedish Yorkshire gilts were used as test animals. They were housed indoors in a conventional stable and tested for signs of standing oestrus, in the presence of a fertile boar, by experienced personnel. Each animal was used only once. At least 1 week prior to initiating the study, the gilts were equipped with permanent jugular vein catheters (*Rodriguez & Kunavongkrit* 1983) so that blood samples could be taken without disturbing the animal. Special care was taken to accustom the gilts to human handling, in order to avoid stress during the test period.

The gilts were either in procestrus or on the first day of cestrus during the test periods.

# Experimental Design

Two ultra-miniature pressure sensors (PR-249, Millar Instr. USA), placed at 2 different points along the same isthmus of the oviduct were used for monitoring intraluminal pressure variations (*Henriksson et al.* 1987). The atmospheric pressure on the day of operation was used as a reference value for the entire test period. Intraluminal pressure recordings were started 9:00 a.m. on the following morning. After an initial 60 min period of intraluminal pressure recordings, the gilts were given an intravenous injection of FM (Finadyne<sup>®</sup>, Schering Corporation, Kenilworth, USA) at the dose of 2.2 mg/kg body weight. The intraluminal pressure recordings were discontin-

ued for 60 min following the FM treatment after which the recordings were resumed and continued for 60 min. Up to this point, blood samples were collected every 30 min. The animals were then injected with prostaglandin  $F_{2\alpha}$  (PGF<sub>2 $\alpha$ </sub>) (Dinolytic<sup>®</sup>, Upjohn Limited, Crawley, England) intravenously, twice, with a 60 min interval separating the injections. Intraluminal pressure was recorded continuously until 60 min had elapsed after the last injection. Gilts nos. 2 and 3 were first treated with 0.1 mg and 60 min later, with 0.5 mg PGF2\_, while gilt no. 1 was treated first with the higher dose and 60 min later with the lower dose. Blood samples were collected at 5 and 10 min after receiving  $PGF_{2\alpha}$  and then every 10 min until 60 min had elapsed from the administration of  $PGF_{2\alpha}$ . Prior to removing the pressure sensors, it was confirmed that the pressure sensors had retained their proper positions. Sections were removed from both oviducts from each gilt for histological examination under a light microscope.

#### **Blood Samples**

All blood samples were collected into heparinized Vacutainer<sup>®</sup> tubes (Becton and Dickinson, USA) and immediately centrifuged. Plasma was removed and stored at -20°C until analysed by radioimmunoassay for concentrations of progesterone (*Bosu et al.* 1976), oestradiol-17ß (*Boilert et al.* 1973) and 15-ketodihydroprostaglandin  $F_{2\alpha}$  (*Granström & Kindahl* 1982). The methods used have earlier been validated for the porcine species (*Kunavongkrit at al.* 1983).

# Calculations

When calculating the results, all calculations were based on the recordings obtained from the distal pressure sensor. The proximal pressure sensor was used for determining the propagation direction of outburst of increased



Fig. 1. Example of the phasic pressure fluctuations in the isthmus. Rp: atmospheric pressure, bp: base pressure, pf: phasic pressure fluctuations.

intraluminal pressure. The base line was set equivalent to the atmospheric pressure of the operation day. Intraluminal pressure could be described as being composed of a base pressure upon which phasic pressure fluctuations were superimposed (Fig. 1). The phasic pressure fluctuations could be arranged in a wavy, irregular or regular pattern with stable resting pressures and amplitudes. Outbursts of increased pressure were defined as marked increases in intraluminal pressure, where the lowest resting pressure of the phasic pressure fluctuations at the peak of the outburst was greater than the total pressure of the registration period. The base pressure was defined as equivalent to the lowest resting pressure during a 5 or 10 min period. The total pressure was derived by adding the mean amplitude of the phasic pressure fluctuations determined for 2 nonconsecutive min when the gilt was relatively still, to the base pressure. The frequency of the pressure fluctuations was also determined. The base pressure, the total pressure and the frequency of the phasic pressure fluctuations were determined for 10 min periods and two 5 min periods after each  $PGF_{2\alpha}$ injection, in connection with each blood sample.

#### Results

Flunixin treatment had a clear inhibitory effect on the circulating levels of 15-ketodihydro-PGF<sub>2α</sub> (Table 2). In all 3 gilts, the frequencies of the phasic pressure fluctuations decreased after treatment with FM. The total pressure recorded in gilt no. 3 was also lower after FM treatment (Table 1). The patterns in which the phasic pressure fluctuations were arranged did not change after FM treatment. No outburst of increased pressure occurred prior to or after FM treatment.

Both injections of  $PGF_{2\alpha}$  resulted in a large increase in the prostaglandin metabolite concentrations (Table 2).

Treatment with  $PGF_{2\alpha}$  resulted in an increase in the frequency of the phasic pressure fluctuations. When the high dose of  $PGF_{2\alpha}$  was used, a transient dramatic effect on the base pressure and the total pressure in the oviductal isthmus was seen (see Table 2). This increase in intraluminal pressure was recorded simultaneously by both pressure sensors.

No adhesions were observed when inspecting the oviducts after completion of the pressure recordings. The pressure sensors had retained their proper positions. No differences were seen when histological preparations from both oviducts of each animal were compared under a light microscope.

# Discussion

The results from this study indicate that endogenous prostaglandins are involved in maintaining the intraluminal pressure in the oviduct during procestrus and day 1 of the oestrous cycle.

Exogenous administration of drugs often results in unphysiological drug tissue levels. The observed effects might therefore be a result of the dose used, rather than of the drug's physiological effect. By studying the effect of a de-

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Table j base pre restraine	l. The ssure ed gilts	effec (B) in SD i	t of i.v. 1 mmH, is the st	admin g, total tandard	uistrati press 1 deviz	ion of ure (T ation o	2.2 mg/kg bod P) in mmHg <i>i</i> f the amplituc	ly weiε and fre les, ex]	tht of quent	flunix cy (F) cd in n	in me of int 1mHg.	glumi ralum , of the	ne (FN inal pi e phas	M) and later ressure fluctu sic pressure fl	admini lations uctuati	stratio in the ions in	n of p isthm cludec	rostag us of 1 1 in ea	landir the ov ch tot	$1 F_{2\alpha}$ c iduct	on the in un- ssure.
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В	752	<i>TT</i> 2	772		762	762	752	822	782	782	<i>7</i> 72	762	752	762	762	<i>7</i> 72	762	752	752	752	762
(SD)	844 (15)	842 (19)	841 (18)		824 (13)	842 (21)	811 (14)	872 (12)	880 (29)	870 (24)	847 (18)	840 (23)	833 (12)	828 (14)	841 (26)	848 (23)	845 (11)	833 (19)	830 (25)	819 (23)	846 (17)
F	36	36	37		31	31	31	49	40	36	34	34	30	32	41	36	32	35	33	32	34
Gilt 2				FM			$PGF_{2\alpha} (0.1 \downarrow)$	l mg)						$PGF_{2\alpha} (0.1)$	5 mg)						
В	750	750	750		760	760	760	770	760	750	750	750	750	750	800	790	780	790	780	780	200
TT (CS)	877 (27)	863 (28)	870 (20)		832 (26)	796 (9)	850 (22)	869 (26)	845 (17)	838 (24)	855 (16)	825 (9)	853 (18)	833 (29)	950 (15)	932 (38)	968 (32)	895 (26)	910 (21)	921 (19)	905 (26)
ц	27	26	25		19	18	17	28	25	22	20	19	20	19	28	23	26	24	22	23	23
Gilt 3				FM			$PGF_{2\alpha} (0.1 \downarrow)$	l mg)						$PGF_{2\alpha} (0.1)$	õ mg)						
В	785	785	775		765	775	765	765	765	765	765	765	765	765	795	805	775	765	775	765	765
TP (SD)	888 (20)	868 (15)	879 (10)		820 (15)	807 (10)	803 (16)	815 (8)	810 (10)	797 (12)	797 (14)	799 (20)	797 (18)	797 (16)	867 (20)	856 (13)	829 (10)	811 (11)	830	(9)	801 (6)
ц	38	36	34		17	12	10	28	23	19	18	20	19	17	47	41	35	27	22	20	2

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			FM ↓				GF <sub>2α</sub> (0.5 m ↓	(g)					PGF	2 <sub>a</sub> (0.1 mg)						
1.8	1.5	2.0	1.7	2.1	2.3	2.4	1.8	0	1.7	2.2	2.7	1.9	2.0	¢ 2.6	2.5	2.6	2.4	2.6	2.1	0.2
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0	1.5	0	2.9	2.1	2.1	2.1	0	1.6	0	1.0	2.2	1.9	0.7	¢ 2.1	2.5	4.4	31	2.5	4.1	3.0
49	52	34	56	39	47	51	60	52	55	59	60	61	58	64	53	51	5.1	59	59	53
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			FM→			д	GF <sub>2α</sub> (0.1 m ↓	(g)					PGF	را (0.5 mg)						
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# Effects of inhibition of prostaglandin biosynthesis

creased endogenous production of prostaglandins on the intraluminal pressure in the oviductal isthmus, possible pharmacological effects of PGF<sub>2 $\alpha$ </sub> could be avoided.

Flunixin has been shown to be a potent inhibitor of prostaglandin synthesis in the porcine species (Odensvik et al. 1989). In the present study, 15-ketodihydro-PGF<sub>2a</sub> levels in peripheral circulation had decreased within 30 min after FM treatment. This is in line with the results obtained by Odensvik et al. (1989), who found that 15-ketodihydro-PGF<sub>2 $\alpha$ </sub> levels in peripheral circulation decreased within 20 min after FM treatment. Further, low levels of 15-ketodihydro-PGF $_{2\alpha}$  were maintained for 5-6 h (Odensvik et al. 1989). It is likely that FM treatment affects the biosynthesis of prostaglandins in all tissues, including that which occurs in the oviduct. It has been shown that  $PGF_{2\alpha}$  when administered both in vitro and in vivo in anaesthetized gilts stimulates oviductal contractility (Rodriguez-Martinez & Einarsson 1985). Furthermore, the decrease in the frequency of the phasic pressure fluctuations seen after FM treatment in the present study, indicates that the endogenous prostaglandin biosynthesis is involved in maintaining the intraluminal pressure in the isthmus of the oviduct during the period studied. During the periovulatory period, when intraluminal pressure in the oviductal isthmus is high (Pettersson 1991), high levels of  $PGF_{2\alpha}$  have been found in the porcine oviductal fluid (Rodriguez-Martinez et al. 1983). The prostaglandins produced in the oviduct may have a stimulating effect on the myosalpinx during this period (Rodriguez-Martinez et al. 1983). Recognizing the need for further studies, it is possible that the decrease in the total pressure seen in the isthmus on day 1 (gilt no. 3) may be a result of decreased levels of prostaglandins in the oviductal fluid. The lack of dramatic effects on the base and total pressures after FM treatment in gilts nos. 1 and 2 was not surprising since other factors, including the rich adrenergic innervation of the isthmus (*Rodriguez-Martinez et al.* 1982 c), are involved in maintaining oviductal intraluminal pressure. In the present study, treatment with PGF<sub>2α</sub> consistently increased the frequency of the phasic pressure fluctuations in an apparantly dose-dependant manner. The higher dose of 0.5 mg also led to a marked increase in the base and total pressures. It would seem that 0.1 mg PGF<sub>2α</sub> had a more physiological effect on the oviduct while a cramp-like condition was induced when 0.5 mg was administered.

# Conclusion

Flunixin meglumine treatment resulted in a marked decrease in circulating levels of 15-ketodihydro-PGF<sub>2α</sub>. The decrease in endogenous prostaglandins resulted in a decreased frequency of the phasic pressure fluctuations in the isthmus of the porcine oviduct. Exogenous administration of PGF<sub>2α</sub> had the reverse effect. The results from this study support the theory that the endogenous prostaglandins are involved in maintaining the intraluminal pressure in the porcine isthmus of the oviduct during prooestrus and oestrus.

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#### References

- Boilert B, Edqvist L-E, Johansson EDB, Lindberg P, Martinsson K: The influence of conjugated estrogens in radioimmunoassays using different antibodies against estradiol-17B. Steriods 1973, 22, 891-894.
- Bosu WTK, Edqvist L-E, Lindberg P, Martinsson K, Johansson EDB: The effect of various dosages of

lynesterol on the plasma levels of oestrogens and progesterone during the menstrual cycle in the rhesus monkey. Contraception 1976, *13*, 677-684.

- *Granström E, Kindahl H:* Radioimmunoassay of the major plasma metabolite of  $PGF_{2\alpha}$ , 15-keto-13,14-dihydro-PGF<sub>2 $\alpha$ </sub>. Meth. Enzymol. 1982, 86, 320-339.
- Henriksson A, Gustavsson A, Einarsson S: A new method for continuous recording of oviductal pressure variations in unrestrained gilts. Acta physiol. scand. 1987, 131, 303-307.
- Kunavongkrit A, Kindahl H, Madej A: Clinical and endocrinological studies in primiparous zeroweaned sows: 2. Hormonal patterns of normal cycling sows after zero-weaning. Zbl. Vet. Med. A 1983, 30, 616-624.
- Odensvik K, Cort N, Basu S, Kindahl H: Effect of flunixin meglumine on prostaglandin  $F_{2\alpha}$  synthesis and metabolism in the pig. J. vet. Pharmacol. Therap. 1989, *12*, 307-311.
- Pettersson A: Cyclic variations in intraluminal pressure in the isthmus of the oviduct in unrestrained gilts. J. vet. Med. A 1991, 38, 337-343.
- *Rodriguez-Martinez, H, Einarsson S:* Influence of prostaglandins on the spontaneous motility of the pig oviducts. Anim. Reprod. Sci. 1985, *8*, 259-279.
- Rodriguez-Martinez H, Einarsson S, Larsson B, Akusu M, Settergren I: Spontaneous motility of the pig oviduct in vitro. Biol. Reprod. 1982a, 26, 98-104.
- Rodriguez-Martinez H, Einarsson S, Larsson B: Spontaneous motility of the pig oviduct in the anaesthetized pig. J. Reprod. Fert. 1982b, 66, 615-624.
- Rodriguez-Martinez H, Garcia B, Ohanian C, Ei-

narsson S: Histochemical investigation on the distribution of adrenergic nerve terminals in the porcine oviduct. Zbl. Vet. Med. A 1982c, 29, 64-71.

- Rodriguez H, Kunavongkrit A: Chronical venous catheterization for frequent blood sampling in unrestrained pigs. Acta vet. scand. 1983, 24, 318-320.
- Rodriguez-Martinez H, Petroni A, Einarsson S, Kindahl H: Concentrations of prostaglandin  $F_{2\alpha}$  in the pig oviductal fluid. Prostaglandins 1983, 25, 413-424.

#### Sammanfattning

Effekten av prostaglandinsynteshämning med flunixin meglumin med efterföljande administrering av prostaglandin  $F_{2\alpha}p^{a}$  intraluminella tryckvariationer i istmusdelen av äggledaren hos gris.

Tre gyltor utrustades med 2 tryckmätare placerade på 2 olika punkter längs samma äggledares isthmus. Efter en inledande period med mätningar av det intraluminella trycket behandlades gyltorna med 2,2 mg flunixin meglumine (FM) per kg kroppsvikt. Blodplasmanivåerna av 15-ketodihydro-PGF $_{2\alpha}$  i perifera cirkulationen sjönk inom 30 minuter efter FM behandlingen. Frekvensen av de fasiska tryckförändringarna sjönk likaså efter FM behandlingen. En kraftig stegring av blodplasmanivåerna av 15-ketodihydro-PG<sub>2 $\alpha$ </sub> påvisades efter exogen tillförsel av PGF2\_. Behandlingen med 0,1 mg PGF<sub>2a</sub> resulterade i en ökad frekvens fasiska tryckfluktationer, medan 0,5 mg PGF<sub>2 $\alpha$ </sub> resulterade i en ökning av såväl basala trycket, totala trycket som av frekvensen fasiska tryckfluktationer.

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