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Brief communication

Mycoplasma alkalescens demonstrated in bronchoalveolar lavage of cattle in Denmark

Branko Kokotovic*, Niels F Friis and Peter Ahrens



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Abstract

Mycoplasma alkalescens is an arginine-metabolizing mycoplasma, which has been found in association with mastitis and arthritis in cattle. Routine bacteriological examination of 17 bronchoalveolar lavage samples from calves with pneumonia in a single herd in Denmark, identified *M. alkalescens* in eight samples. The organism was found as a sole bacterilogical findings in five of the samples as well as in combination with *Mannheimia haemolytica*, *Haemophilus somni* and *Salmonella* Dublin. This is the first report of isolation of *M. alkalescens* in Denmark.

Findings

Mycoplasma alkalescens is a bovine mycoplasma species, which was originally isolated from the nasal cavity of cattle in Australia [1]. Like many other mycoplasmas, M. alkalescens is a normal inhabitant of the upper respiratory tract, but it has also been associated with disease. M. alkalescens has mostly been implicated in mastitis in cattle. It has been isolated from bulk tank milk samples, as well as from outbreaks and sporadic cases of clinical mastitis [2-5]. Furthermore, M. alkalescens has been isolated from cases of severe arthritis, and its ability to induce joint lesions has been confirmed under experimental conditions [6,7]. Rosenfeld & Hill [8] isolated M. alkalescens in pure culture from abomasum and lung of an aborted bovine foetus, while Lamm et al. [9] found M. alkalescens in association with otitis in calves. Finally, M. alkalescens has occasionally been found in association with disorders of the respiratory tract [10,11].

Recently, 17 bronchoalveolar lavage samples from calves suffering from pneumonia in a single herd in Denmark were submitted, on two occasions, to the National Veterinary Institute for laboratory examinations. The samples were examined for the presence of bacterial pathogens, bovine respiratory mycoplasmas, as well as for the presence of bovine respiratory syncytial virus (BRSV), bovine coronavirus and parainfluenza-3 virus (PI-3). The samples were not examined for infectious bovine rhinotracheitis virus, as Denmark is considered free of this infection. Thereby, eight arginine-metabolizing mycoplasmas were isolated. In the present study the identification of the isolates as *M. alkalescens* is presented. This is the first report of isolation of this species in Denmark.

Bacteriological examination was performed according to standard laboratory procedures. Examination for BRSV, bovine coronavirus and PI-3 was performed using an indirect sandwich-ELISA assay [12]. Isolation of mycoplasmas was performed according to standard laboratory procedures using a Hayflick's type of medium enriched with arginine [13], with and without addition of 5% of rabbit hyperimmune antiserum against *Mycoplasma bovirhinis*. The isolates were filtered through 0.45 µm membrane filters (Millipore), cloned and submitted to serologic identification, which was performed by the disc growth inhibition test (DGI) with 6 mm filter paper discs prepared with rabbit hyperimmune antisera against *M. alkalescens* PG51^T (ATCC 29103; NCTC 10135), *Mycoplasma arginini* G230^T (ATCC 23838; NCTC 10129) and *Mycoplasma canadense* 275C^T (ATCC 29418; NCTC 10152), as well as by the indirect epi-immunofluorescence test (IF) on colonies on solid medium. Molecular identification of the isolates was performed by amplification and sequencing of their 16S rRNA genes by using universal primers [14].

Laboratory examinations revealed the presence of several bacterial pathogens in 15 of the 17 bronchoalveolar lavage samples, while no pathogenic bacteria were detected in the two remaining samples. Pasteurella multocida, Mannheimia haemolytica, Histophilus somni and Salmonella Dublin were found in three, three, one and one samples, respectively. Mycoplasma bovirhinis was found in five samples, while arginin-metabolizing mycoplasmas were found in eight samples. Two P. multocida, three M. bovirhinis and five arginin-metabolizing mycoplasma isolates were found as sole bacteriological findings in 10 respective samples. From five samples multiple bacterial species were isolated: M. bovirhinis was found once in combination with P. multocida and once in combination with M. haemolytica, while arginin-metabolizing mycoplasmas were found once in combination with M. haemo*lytica*, once in combination with S. Dublin and once in combination with M. haemolytica and H. somni. No viral pathogens were detected in any of the analysed samples.

Further examinations on arginin-metabolizing mycoplasmas showed that all eight isolates could be serologically identified as *M. alkalescens* and that they were clearly different from the two other arginine-degrading species that are commonly found in cattle, *M. canadense* and *M. arginini* (Table 1). The reactions were clear-cut for the DGI tests showing approximately 3–5 mm broad zones of total or nearly total inhibition around the disc. In the IF test some minor cross reactions were noted when three of the isolates and the type strain of *M. alkalescens* PG51^T were tested with anti-*M. canadense* 275C^T hyperimmune serum. The 16S rDNA sequences of the analysed isolates were identical to each other and to the 16S rDNA sequence of the type strain *M. alkalescens* PG51^T (GenBank accession no. <u>U44764</u>; [15]), which corroborated the serological identification of the isolates as *M. alkalescens*.

Previous studies have shown that H. somni, M. haemolytica and P. multocida are the bacteria that are most commonly associated with bronchopneumonia in calves in Denmark, although they are also part of the normal bacterial flora of the respiratory tract [16,17]. Also, S. Dublin, which is predominantly found in cattle, is capable of causing pneumonia in calves. Isolation of these bacterial species in the submitted samples most likely indicate their role as the primary cause of respiratory disease in the herd, probably in combination with unidentified environmental factors. In addition to the bacteria, M. bovirhinis and M. alkalescens were also isolated. M. bovirhinis is commonly found as a part of the normal respiratory tract microflora of cattle and is not considered to be pathogenic. M. alkalescens, however, has been found in association with disorders of the respiratory tract but its role as a respiratory pathogen remains equivocal. Experimental infections have demonstrated that M. alkalescens has the ability to colonize lung tissue [18], but the two strains used in the study apparently failed to produce pneumonia [18]. In the present study, we found M. alkalescens in pure culture

 Table I: Serological identification of eight mycoplasma field isolates from cattle by DGI and IF test using type strain antisera against

 Mycoplasma alkalescens, Mycoplasma arginini and Mycoplasma canadense.

Mycoplasma strains	Antiserum					
	M. alkalescens PG51 [⊤]		M. arginini G230 [⊤]		M. canadense 275C [⊤]	
	DGlª	ΙF ^ь	DGlª	IF ^b	DGlª	IF⁵
M. alkalescens PG51 [⊤]	5	+	0	-	0	?
Field strain MK19/02	≥ 3	+	0	-	0	-
Field strain MK20/02	≥ 3	+	0	-	0	?
Field strain MK21/02	≥ 3	+	0	-	0	-
Field strain MK49/02*	≥ 3	+	0	-	0	?
Field strain MK51/02	≥ 3	+	0	-	0	-
Field strain MK52/02*	≥ 3	+	0	-	0	-
Field strain MK53/02	≥ 3	+	0	-	0	-
Field strain MK56/02*	≥ 3	+	0	-	0	?
M. arginini G230 [⊤]	0	-	5	+	0	-
M. canadense 275C [⊤]	0	-	0	-	5	+

^a zone of inhibition in mm; ^b distinct FITC colour of stained colonies; ? inconclusive results; * examined by 16S rDNA sequencing.

in five of the analysed bronchoalveolar lavage samples. This finding is, however, not sufficient to warrant a role of M. alkalescens as a cause of bronchopneumonia, since we also found two samples containing only a non-pathogenic M. bovirhinis, and two samples without any bacterial or viral pathogens, despite the fact that they derived from calves with clinical signs of a respiratory disease. The failure to detect respiratory tract pathogens in these samples may be due to i.e., their absence in a particular disease stadium when sampling took a place or due to antibiotic treatment. Taking into the consideration the overall bacteriological findings of this study, it seems likely that M. alkalescens may have had a role either as a secondary invader or as an opportunistic pathogen rather than suggesting a causal role of the organism in the pneumonia complex.

M. alkalescens is regarded as one of the most common causative agents of mastitis [19]. So, with demonstration of the organism in a Danish cattle herd, a further member of *Mycoplasma* is added to the group of bovine mastitis-inducing microorganisms in Denmark. The only mycoplasma species isolated from clinical outbreaks of mastitis in Denmark so far has been *Mycoplasma bovis* [20], while other mastitis-inducing species, *Mycoplasma bovigenitalium* and *M. canadense*, have been isolated only from the respiratory and the genital tract and semen samples [21]. Further investigations are needed in order to determine the prevalence of *M. alkalescens* in the Danish cattle population and, indeed, to draw a firm conclusions on its importance in disease conditions other than mastitis.

Competing interests

The author(s) declare that they have no competing interests.

Authors' contributions

BK carried out bacteriological examination of samples, isolation and cloning of mycoplasmas and drafted the manuscript.

NFF carried out identification of mycoplasma isolates by DGI test and epi-immunofluorescence.

PA carried out molecular identification of mycoplasma isolates.

All authors read and approved the final manuscript.

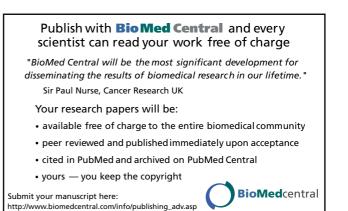
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References

 Hudson JR, Etheridge JR: A new type of pleuropneumonia-like microorganism (PPLO) from the nose of cattle. Aust Vet J 1963, 39:1-5.

- 2. Brookbanks EO, Carter ME, Holland JTS: Mycoplasma mastitis. NZ Vet / 1969, 17:179-180.
- 3. Dellinger JD, Jasper DE, Ilic M: Characterization studies on mycoplasmas isolated from bovine mastitis and the bovine respiratory tract. *Cornell Vet* 1977, **67**:351-360.
- 4. Jasper DE: Prevalence of mycoplasmal mastitis in the Western States. *California Vet* 1980, 34:24-26.
- 5. Jasper DE: The role of Mycoplasma in bovine mastitis. J Amer Vet Med Assoc 1982, 181:158-162.
- 6. Bennett RH, Jasper DE: Mycoplasma alkalescens-induced arthritis in dairy calves. J Amer Vet Med Assoc 1978, 172:484-488.
- 7. Whithear KG: Isolation of Mycoplasma alkalescens from cases of polyarthritis in embryo-transplant calves. Aust Vet J 1983, 60:191-192.
- 8. Rosenfeld LE, Hill MWM: The isolation of Mycoplasma alkalescens from an aborted bovine foetus. Aust Vet J 1980, 56:350.
- Lamm CG, Munson L, Thurmond MC, Barr BC, George LW: Mycoplasma otitis in California calves. J Vet Diagn Invest 2004, 16:397-402.
- Hewicker-Trautwein M, Feldmann M, Kehler W, Schmidt R, Thiede S, Seeliger F, Wohlsein P, Ball HJ, Buchenau I, Spergser J, Rosengarten R: Outbreak of pneumonia and arthritis in beef calves associated with Mycoplasma bovis and Mycoplasma californicum. Vet Rec 2002, 151:699-703.
- 11. Ayling RD, Bashiruddin SE, Nicholas RAJ: Mycoplasma species and related organisms isolated from ruminants in Britain between 1990 and 2000. Vet Rec 2004, 155:413-416.
- Uttenthal A, Jensen NPB, Blom JY: Viral aetiology of enzootic pneumonia in Danish dairy herds: Diagnostic tools and epidemiology. Vet Rec 1996, 139:114-117.
- 13. Kobisch M, Friis NF: Swine mycoplasmosis. Rev Sci Tech Off Int Epiz 1996, 15:1569-1605.
- Weisburg WG, Barns SM, Pelletier DA, Lane DJ: 16S ribosomal DNA amplification for phylogenetic study. J Bacteriol 1991, 173:697-703.
- Pettersson B, Uhlen M, Johansson K-E: Phylogeny of some mycoplasmas from ruminants based on 16S rRNA sequences and definition of a new cluster within the hominis group. Int J Syst Bacteriol 1996, 46:1093-1098.
- Tegtmeier C, Uttenthal A, Friis NF, Jensen NE, Jensen HE: Pathological and microbiological studies on pneumonic lungs from Danish calves. J Vet Med 1999, 46:693-700.
 Watts JL, Yancey RJ, Sarah JR, Salmon SA, Case CA: A 4-year survey
- Watts JL, Yancey RJ, Sarah JR, Salmon SA, Case CA: A 4-year survey of antimicrobial susceptibility trends for isolates from cattle with bovine respiratory disease in North America. J Clin Microbiol 1994, 32:725-731.
- Gourlay RN, Howard CJ, Thomas LH, Wyld SG: Pathogenicity of some Mycoplasma and Acholeplasma species in the lungs of gnotobiotic calves. Res Vet Sci 1979, 27:233-237.
- 19. Gonzalez RN, Wilson DJ: Mycoplasma mastitis in dairy herds. Vet Clin North Am Food Anim Pract 2003, 19:199-221.
- Friis NF: Mycoplasma bovis-induced mastitis in cattle in Denmark. Nord Vet Med 1984, 36:324-325.
- 21. Friis NF, Blom E: Isolation of Mycoplasma canadense from bull semen. Acta Vet Scand 1983, 24:315-317.



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