

Research

Open Access

Prevalence of *Borrelia burgdorferi* sensu lato and *Anaplasma phagocytophilum* in questing *Ixodes ricinus* ticks in relation to the density of wild cervids

Olav Rosef*¹, Algimantas Paulauskas² and Jana Radzijeuskaja²

Address: ¹Telemark University College, Bø i Telemark, Norway and ²Vytautas Magnus University, Kaunas, Lithuania

Email: Olav Rosef* - olav.rosef@hit.no; Algimantas Paulauskas - a.paulauskas@gmf.vdu.lt; Jana Radzijeuskaja - j.radzijeuskaja@bs.vdu.lt

* Corresponding author

Published: 27 November 2009

Received: 28 April 2009

Acta Veterinaria Scandinavica 2009, 51:47 doi:10.1186/1751-0147-51-47

Accepted: 27 November 2009

This article is available from: <http://www.actavetscand.com/content/51/1/47>

© 2009 Rosef et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: *Borrelia burgdorferi* sensu lato and *Anaplasma phagocytophilum* have been considered as pathogens in animals and humans. The role of wild cervids in the epidemiology is not clear. We analyzed questing *Ixodes ricinus* ticks collected in spring for these pathogens from sites with high (Fjelløyvær and Strøm) and low density (Tjore, Hinnebu and Jomfruland) of wild cervids to study the spread of the pathogens in questing ticks.

Methods: For detection of *Anaplasma phagocytophilum* a 77-bp fragment in the *msp2* gene was used. Detection of *Borrelia burgdorferi* sensu lato was performed using the FL6 and FL7 primers according to sequences of conserved regions of the *fla* gene. The *OspA* gene located on the linear 49-kb plasmid was used as target in multiplex PCR for genotyping. Genospecies-specific primers were used in the PCR for *Borrelia burgdorferi* sensu stricto, *B. afzelii* and *B. garinii*.

Results: Infection rates with *Borrelia* spp. were significantly lower at Fjelløyvær and Strøm compared to Tjore and Hinnebu; Fjelløyvær vs. Tjore ($\chi^2 = 20.27$, $p < 0.0001$); Fjelløyvær vs. Hinnebu ($\chi^2 = 24.04$, $p < 0.0001$); Strøm vs. Tjore ($\chi^2 = 11.47$, $p = 0.0007$) and Strøm vs. Hinnebu ($\chi^2 = 16.63$, $p < 0.0001$). The *Borrelia* genospecies were dominated by *B. afzelii* (82%) followed by *B. garinii* (9.7%) and *B. burgdorferi* sensu stricto (6.9%). *B. burgdorferi* s.s. was only found on the island of Jomfruland. The infection rate of *Anaplasma phagocytophilum* showed the following figures; Fjelløyvær vs. Hinnebu ($\chi^2 = 16.27$, $p = 0.0001$); Strøm vs. Tjore ($\chi^2 = 13.16$, $p = 0.0003$); Strøm vs. Hinnebu ($\chi^2 = 34.71$, $p < 0.0001$); Fjelløyvær vs. Tjore ($\chi^2 = 3.19$, $p = 0.0742$) and Fjelløyvær vs. Støm ($\chi^2 = 5.06$, $p = 0.0245$). Wild cervids may serve as a reservoir for *A. phagocytophilum*. Jomfruland, with no wild cervids but high levels of migrating birds and rodents, harboured both *B. burgdorferi* s.l. and *A. phagocytophilum* in questing *I. ricinus* ticks. Birds and rodents may play an important role in maintaining the pathogens on Jomfruland.

Conclusion: The high abundance of roe deer and red deer on the Norwegian islands of Fjelløyvær and Strøm may reduce the infection rate of *Borrelia burgdorferi* sensu lato in host seeking *Ixodes ricinus*, in contrast to mainland sites at Hinnebu and Tjore with moderate abundance of wild cervids. The infection rate of *Anaplasma phagocytophilum* showed the opposite result with a high prevalence in questing ticks in localities with a high density of wild cervids compared to localities with lower density.

Background

Lyme disease, an important arthropod-borne disease of humans in the northern hemisphere, can manifest in many organ systems with symptoms including skin rashes, meningitis, optic neuritis, facial nerve palsy and atrioventricular nodal block. Failure to treat infection promptly and adequately can result in long-term debilitating effect on the patient's health. Three species have been proven to be pathogenic in humans: *Borrelia afzelii*, *B. garinii* and *B. burgdorferi* sensu stricto [1]. These species appear to be responsible for causing different clinical syndromes [2].

It is well known that *Ixodes* ticks feed on deer species [3], and that high abundance of *Ixodes* ticks follows a high abundance of deer [4], but the role of cervid species in the epidemiology of Lyme disease is not completely understood. Although it has been suggested that adaptive immune responses may be involved in the regulation of spirochete transmission [5], the detailed mechanisms underlying differential transmission of the *Borrelia* genospecies by hosts are unknown. Investigators have concluded that roe deer (*Capreolus capreolus*) [6] and red deer (*Cervus elaphus*) [7-9] are incompetent reservoirs for *B. burgdorferi*. Spirochaetes that are sensitive to destruction by the complement system of a particular host species are lysed early in the midgut of the feeding tick and are thereby eliminated by the host [10]. These findings have led to the hypothesis that the host range of spirochaete strain is restricted by its repertoire of genes that encode ligands with the high binding affinities for complement inhibition [7].

Tick-borne fever caused by *A. phagocytophilum* has been considered a common disease in domestic ruminants along the coast of southern Norway [11]. Several other mammalian species including wild cervids have also been found infected with *A. phagocytophilum* [12]. Stuen et al. [13] found seroprevalences of granulocytic *Ehrlichia* spp. in moose (*Alces alces*) of 43%, red deer 55%, and roe deer 96% from *I. ricinus* infested counties in Norway. A study in Switzerland found serological evidence of granulocytic ehrlichial infection in roe deer [14].

Human granulocytic anaplasmosis (HGA) caused by *Anaplasma phagocytophilum* was first identified in 1990 in a patient who died [15]. HGA is increasingly recognized as an important and frequent cause of fever after tick bite world wide [16], including Scandinavia [17] where *Ixodes* ticks bite humans. Several *Ixodes* spp. including *I. ricinus*, *I. arboricola*, *I. calidonicus*, *I. frontalis*, *I. hexagonus*, *I. lividus*, *I. persulcatus*, *I. trianguliceps*, *I. urinae* and *I. unicavatus* have been found in Fennoscandia [18-20] Most human cases occur between June and August and usually appear as an

undifferentiated febrile illness. The incubation period following tick-bite is 7-10 days and symptoms include high fever, rigors, generalized myalgias, severe headache and malaise [16]. Bjöersdorff et al. [17] found a seroprevalence of 15-20% among 1000 tick-exposed patients (mainly from Sweden and Norway) and concluded a widespread exposure to granulocytic *Ehrlichia* (now *Anaplasma* spp.). In Slovenia 3.2% of *I. ricinus* were infected with *Anaplasma*, and they were 99.8% identical to those previously determined from human patients [21]. The main vector in Europe is *I. ricinus*. In other continents Zhang et al. [22] found a high seroprevalence rate (8.8%) for *A. phagocytophilum* among 365 farm-workers in China and suggested that human infections with these zoonotic bacteria are frequent and largely unrecognized. A seroprevalence between 2.3% and 5.6% was found in different locations in Mongolia and Walder et al. [23] concluded that *A. phagocytophilum* is endemic. Brown et al. [24] confirmed that woodland rodents can maintain *A. phagocytophilum* in Great Britain in the absence of other reservoir hosts which suggests that *I. trianguliceps* is a competent vector.

The aim of the present study was to compare the prevalence of *B. burgdorferi* s.l. and *A. phagocytophilum* in *I. ricinus* ticks in sites with both high and low abundance of roe deer, red deer and moose to evaluate the role of wild cervids in the epidemiology.

Materials and methods

Locations and habitats

Tick samples were collected on two islands on the coast of western Norway: at Strøm (N7048360E498426), on the island of Hitra, and on the island of Fjelløyvær (N7059209E504490) close to the main island Hitra and connected by a bridge. Both islands are largely covered with heath and a mixture of deciduous and pine forest. There are no foxes, but sea gulls and raptorial birds are common, and roe deer and red deer densities are high. There are farms on both islands with grass production and grazing cattle and sheep. Tick samples were also collected at three sites along the southern coast of Norway. These included Tjore, a coastal mainland site (N6463382E473032) located in a mixture of farmland and mixed deciduous, pine and spruce forest, and within 100 m outside of a red deer enclosure; Hinnebu (N6493848E469418) situated 30 km from the coast with similar mixed forest but no agriculture or grazing domestic animals; and Jomfruland, an island with agriculture and mixed forest (N6524446E533677). Jomfruland is frequented by many migrating birds and is grazed by sheep and cattle, but contains no wild cervids. Coordinates are given in UTM32 (Euref 89) values.

Abundance of roe deer, red deer and moose

We used the official municipal hunting statistics for 2007 for each township involved to estimate the numbers of game animals at each site (Table 1). We have defined low density as less than one animal killed per km² and high density as more than 3.

Tick collection

Questing *I. ricinus* ticks were collected during spring (April-May) 2006-2008 at all five locations using the standard flagging method [25] by drawing a 1 m² piece of cotton cloth over the vegetation. Ticks attached to the towel were picked with tweezers and placed into 1.5 ml test tube filled with 70% ethanol.

DNA extraction and detection of *Ixodes ricinus*

A modified procedure for extracting DNA with ammonium hydroxide solution (2.5%) was performed [26,27]. The lysates were stored at -20°C until use. For *I. ricinus* identification, the lysates were analysed with species-specific primers IxriF and IxriR resulting in a 150 bp segment of the 5.8 srRNA gene [28,29]. This PCR reaction was further used as positive control. DNA bands were stained with ethidium bromide and visualised by UV transillumination (EASY Win32, Herolab, Germany).

Detection of *Borrelia burgdorferi sensu lato*

The occurrence of *Borrelia burgdorferi* s.l. in ticks was determined by polymerase chain reaction by using the oligonucleotide primers FL6 and FL7 according to sequences of conserved regions of the *fla* gene [26]. PCR products were resolved by 1.5% agarose gel electrophoresis with addition of ethidium bromide and visualized under UV light (EASY Win32, Herolab, Germany). The achieved specific amplification products of 276 base pairs (bp) were considered a positive result. Negative and positive controls were included in all runs.

Genotyping of *Borrelia burgdorferi sensu lato*

The *OspA* gene located on the linear 49-kb plasmid was used as target in multiplex PCR according to Demaer-

shalck et al. [30]. Genospecies-specific primers were used in the PCR for *B. burgdorferi sensu stricto*, *B. afzelii* and *B. garinii*. PCR amplification products were resolved onto 2.0% agarose gel electrophoresis and visualized under UV light. The specific products of 544 bp (*B. burgdorferi* s.s.) 345 bp (*B. garinii*) and 189 bp (*B. afzelii*) were considered to represent positive results. Negative and positive controls were included in all runs.

Detection of *Anaplasma phagocytophilum*

I. ricinus questing ticks were examined for the prevalence of *A. phagocytophilum* by using the species-specific primers ApMSP2f, ApMSP2r, and TaqMan probe ApMSP2p-FAM, as described by Courtney et al. [31]. A 77-bp fragment in the *msp2* gene of *A. phagocytophilum* was amplified. PCR was performed using TaqMan Master Mix (Applied Biosystems, CA) in a quantitative thermal cycler (iCycler, Bio-Rad Laboratories, Inc., Hercules, CA). Negative and positive controls were included in all runs.

Statistics

The data were analysed statistically by means of Pearson's χ^2 test by using the statistical package STATISTICA for WINDOWS 5.5. We compared the mean isolation rate of *B. burgdorferi* s.l. and *A. phagocytophilum* for 2006-2008 in sites with different densities of wild cervids.

Results

The highest density of wild cervids was Fjelløyvær followed by Strøm (Table 1). No *Borrelia* was detected in questing ticks in Fjelløyvær, and low values in Strøm during the three year period (Table 2). The infection rates were significantly lower in areas with high density of wild cervids compared to sites with low density: Fjelløyvær vs. Tjore ($\chi^2 = 20.27$, $p < 0.0001$); Fjelløyvær vs. Hinnebu ($\chi^2 = 24.04$, $p < 0.0001$); Strøm vs. Tjore ($\chi^2 = 11.47$, $p = 0.0007$) and Strøm vs. Hinnebu ($\chi^2 = 16.63$, $p < 0.0001$). There were significantly lower values on Fjelløyvær vs. Jomfruland ($\chi^2 = 10.66$, $p = 0.0011$); Fjelløyvær vs Strøm ($\chi^2 = 4.26$, $p = 0.0390$) and Hinnebu vs. Jomfruland ($\chi^2 = 6.56$, $p = 0.0104$), but no significant difference between Tjore vs. Jomfruland ($\chi^2 = 3.2$, $p = 0.0735$); Strøm vs. Jomfruland ($\chi^2 = 3.24$, $p = 0.0719$) and Hinnebu vs. Tjore ($\chi^2 = 0.27$, $p = 0.6006$). The distribution of genospecies is shown in Table 3. *B. afzelii* dominated with 82% followed by *B. garinii* (9.7%) and *B. burgdorferi* s.s. (6.9%). *B. burgdorferi* s.s. was only found on the island of Jomfruland.

The prevalence of *A. phagocytophilum* infections in questing ticks (Table 4) was significantly higher in localities with high density of wild cervids compared to localities with lower density (Table 1): Fjelløyvær vs. Hinnebu ($\chi^2 = 16.27$, $p = 0.0001$); Fjelløyvær vs. Strøm ($\chi^2 = 5.06$, $p = 0.0245$); Strøm vs. Tjore ($\chi^2 = 13.16$, $p = 0.0003$) and Strøm vs. Hinnebu ($\chi^2 = 34.71$, $p = 0.0000$). The figures

Table 1: Number of animals killed by hunting per km² (hunting statistics for 2007)

	Red deer	Roe deer	Moose	Total
Fjelløyvær	0.05 (1) ^a	8.62 (181)	0*	8.67
Strøm	1.94 (846)	1.18 (513)	0*	3.12
Hinnebu	0.05 (30)	0.33 (198)	0.31 (194)	0.69
Tjore	0.02 (6)	0.56 (528)	0.17 (194)	0.75
Jomfruland	0**	0**	0*	0

^aThe numbers in parentheses represent the total number of killed animals.

*Moose is absent.

**Red deer and roe deer are absent.

Table 2: Prevalence of *Borrelia burgdorferi* sensu lato in questing *Ixodes ricinus* ticks 2006, 2007 and 2008.

Locations	Years	Female		Male		Nymph		Total					
		N	Prevalence	N	Prevalence	N	Prevalence	N	Prevalence				
		n	%	n	%	n	%	n	%				
Hitra, Fjelløyvær	2008	9	0	0	8	0	0	43	0	0	60	0	0
	2007	23	0	0	27	0	0	30	0	0	80	0	0
	2006	2	0	0	6	0	0	56	0	0	64	0	0
											204	0	0
Hitra, Strøm	2008	5	0	0	7	1	14.3	40	0	0	52	1	1.9
	2007	40	0	0	32	0	0	16	0	0	88	0	0
	2006	5	1	20	3	0	0	89	3	3.4	97	4	4.1
											237	5	2.1
Tjore	2008	24	1	4.2	14	0	0	20	2	10	58	3	5.2
	2007	19	2	10.5	23	3	13	38	1	2.6	80	6	7.5
	2006	4	1	25	4	0	0	16	7	43.8	24	8	33.3
											162	17	10.5
Hinnebu	2008	31	3	9.7	41	3	7.3	7	0	0	79	6	7.6
	2007	52	15	28.8	57	4	7	6	0	0	115	19	16.5
	2006	42	4	9.5	32	4	12.5	32	4	12.5	106	12	11.3
											300	37	11.8
Jomfruland	2008	29	3	10.3	30	0	0	20	0	0	79	3	3.8
	2007	30	3	10	20	2	10	22	0	0	72	5	6.9
	2006	8	0	0	8	0	0	76	7	9.2	92	5	5.4
											243	13	5.3

N = number of tested ticks; n = number of infected ticks

for Hinnebu vs. Tjore was ($\chi^2 = 5.07$, $p = 0.0243$); Hinnebu vs. Jomfruland ($\chi^2 = 30.73$, $p = 0.000$) and Jomfruland vs. Tjore ($\chi^2 = 10.97$, $p = 0.0009$). There was one exception, with no significant difference between Fjelløyvær and Tjore where a high level of *A. phagocytophilum* was detected in 2008 ($\chi^2 = 3.19$, $p = 0.0742$) (Table 4). There were no significant difference between Strøm and Jomfru-

land ($\chi^2 = 0.38$, $p = 0.54$), or Fjelløyvær and Jomfruland ($\chi^2 = 3.78$, $p = 0.0519$).

Discussion

Kurtenbach et al. [5] showed that sera from red deer were indiscriminating borrelicidal for the three human pathogenic strains. The reservoir incompetence of roe deer [32]

Table 3: *Borrelia burgdorferi* sensu lato genospecies in questing *Ixodes ricinus* ticks.

Locations	<i>B.afzelii</i> n/N (%)	<i>B.garinii</i> n/N (%)	<i>B.burgdorferi</i> s.s. n/N (%)	<i>B.burgdorferi</i> s.s.+ <i>B.afzelii</i> n/N (%)
Hitra, Strøm	4/5 (80)	1/5 (20)	0/5 (0)	0/5 (0)
Tjore	13/17 (76.5)	4/17 (23.5)	0/17 (0)	0/17 (0)
Hinnebu	35/37 (94.6)	2/37 (5.6)	0/37 (0)	0/37 (0)
Jomfruland	7/13 (53.8)	0/13 (0)	5/13 (38.5)	1/13 (7.7)
Total	59/72 (82)	7/72(9.7)	5/72(6.9)	1/72(1.4)

N = number of tested ticks; n = number of infected ticks; (%) - prevalence of infection

Table 4: Prevalence of *Anaplasma phagocytophilum* in questing *Ixodes ricinus* ticks 2006, 2007 and 2008.

Locations	Years	Female		Male			Nymphs		Total				
		N	Prevalence	N	Prevalence		N	Prevalence	N	Prevalence			
		n	%	n	%	%	n	%	n	%			
Hitra, Fjelløyvær	2008	9	0	0	8	0	0	42	2	4.8	59	2	3.4
	2007	23	4	17.4	24	3	12.5	30	2	16.7	77	12	15.6
	2006	2	1	50	6	0	0	56	2	3.6	64	3	4.7
											200	17	8.5
Hitra, Strøm	2008	5	0	0	7	2	28.6	40	11	27.5	52	13	25
	2007	40	9	22.5	35	9	25.7	33	3	9.1	108	21	19.4
	2006	5	1	20	3	1	33.3	89	8	8.9	97	10	10.3
											257	44	17.1
Tjore	2008	24	3	12.5	14	0	0	20	2	0.8	58	5	8.6
	2007	19	0	0	22	0	0	21	0	0	63	0	0
	2006	4	0	0	4	0	0	16	0	0	24	0	0
											145	5	3.4
Hinnebu	2008	31	1	3.2	41	0	0	7	0	0	79	1	1.3
	2007	27	0	0	18	0	0	5	0	0	50	0	0
	2006	42	0	0	32	0	0	32	0	0	106	0	0
											235	1	0.4
Jomfruland	2008	29	6	20.7	40	10	25	57	13	22.4	126	29	23.0
	2007	50	8	16	32	3	9.4	49	4	8.2	131	15	11.5
	2006	8	1	12.5	8	1	12.5	75	6	8	91	8	8.7
											348	52	14.9

N = number of tested ticks; n = number of infected ticks

and red deer [9] correlates with this borrelicidal effect. Complement appears relevant to host incompetency for *Borrelia*, and this carries over to prevent tick infection and lyse the spirochetes early in the midgut of the feeding tick, and are thereby eliminated by the host [10]. Low levels of *B. burgdorferi* s.l. in ticks were found in both sites on Hitra (Table 2). No infected ticks were detected in Fjelløyvær during the three year period, and only a low level of *B. burgdorferi* s.l. in 2006 (4.1%) and in 2008 (1.9%) at Strøm (Table 2). Fjelløyvær has a very high abundance of roe deer, but red deer are nearly absent (Table 1). Strøm has a high abundance of both red and roe deer. We believe that the main route for the tick cycles is red deer and roe deer at Strøm and Fjelløyvær. The high abundance of deer gives high levels of ticks, but the serum incompetence will reduce both the infection in ticks and the risk of Lyme disease transmission.

This contrasts with the figures at Hinnebu where the infection rates with *B. burgdorferi* s.l. were 10%, 16.5% and 7.6% in 2006-2008 (Table 2). Hinnebu is forest-covered with a low density of moose and roe deer, and a low abundance of red deer. Harvest statistics show a much lower combined density of wild cervids at Hinnebu than at Fjelløyvær and Strøm (Table 1). Tjore has low densities of red deer and moose, and a moderate density of roe deer. Ticks collected outside a fenced red deer farm indicated that the presence of the farm had no influence on the level of *B. burgdorferi* s.l. The overall infection rates in ticks were 33.3% in 2006, 6.9% in 2007 and 5.2% in 2008. The capacity of deer to act as reservoirs for *B. burgdorferi* s.l., is controversial [33,34]. However, our results clearly support the idea that wild cervids are incompetent reservoirs. Our results showed that the infection rates in questing ticks were significantly lower in areas with a high density

of wild cervids (Fjelløyvær and Strøm) compared to sites with low density (Tjore and Hinnebu) (Tables 1 and 2).

B. afzelii genospecies from ticks dominated with 82% as shown in Table 3. This genospecies is related to rodents [7,35,36]. *B. garinii* was detected in Strøm, Tjore and Hinnebu while *B. burgdorferi* s.s was found on questing ticks from Jomfruland. Though Jomfruland has no wild cervids, it does have grazing domestic animals, plus migrating birds during spring and autumn. In this site we investigated 49 *A. flavicollis* mice and found an infection rate of 12.2% with *B. burgdorferi* s.l. Of 490 *I. ricinus* ticks feeding on rodents, 17 (3.5%) were infected with *B. burgdorferi* s.l., and *B. burgdorferi* s.l. was also detected in 15.3% (n = 262) of ticks feeding on blackbirds *Turdus merula* [Rosef, unpublished]. It seems that birds and rodents play an important role in maintaining *Borrelia* infection on Jomfruland. The prevalence of *B. burgdorferi* s.l. in ticks showed significantly lower values on Fjelløyvær than Jomfruland and Hinnebu than Jomfruland. In comparison there was no significance between Tjore and Jomfruland and Strøm and Jomfruland.

In contrast to infection with *B. burgdorferi* s.l., cervids are important reservoirs for *A. phagocytophilum*. Stuen et al. [13] found an overall high seroprevalence for *A. phagocytophilum* (formerly granulocytic *Ehrlichia* spp.) in moose, red deer and roe deer in Norway with 43%, 55% and 96% respectively. Experimental *Anaplasma* infection in red deer has shown subclinical persistent infection [37]. These wild ruminants are exposed to *A. phagocytophilum* and comprise the most widespread tick-borne infection in animals in Europe [38]. In Wisconsin, Michalski et al. [39] found a prevalence in ticks between 5.8% and 8.9%, and in white-tailed deer between 11.5% and 26% using PCR and DNA sequencing. A parietic condition in an *A. phagocytophilum* infected roe deer calf [40] and ehrlichiosis in a moose calf [12] has been observed in Norway. The high level of infected ticks at Fjelløyvær and Strøm (Table 4) not surprisingly shows that roe deer and red deer probably are competent reservoirs and vehicles for this bacterium.

A low prevalence of *A. phagocytophilum* in ticks from Hinnebu and Tjore was found in 2008 (Table 4) but it could not be detected in 2006 and 2007. The prevalence of *A. phagocytophilum* in host seeking *I. ricinus* ticks in Norway varied from zero to 19.4% in 18 sites investigated, with the highest prevalence occurring in Hitra [41]. The prevalence of *A. phagocytophilum* infections in ticks was significantly higher in localities with high density of wild cervids (Fjelløyvær and Strøm) compared to localities with lower densities (Tjore and Hinnebu) (Tables 1 and 4). An exception that cannot be explained occurred in 2008 when the

prevalence of *A. phagocytophilum* was high in Tjore and low in Fjelløyvær.

In Europe *B. burgdorferi* s.l. and *A. phagocytophilum* are transmitted by the same vector (*I. ricinus*), but it is unclear whether both pathogens use the same range of host species as reservoirs on a smaller scale. In Europe, studies conducted in the United Kingdom, Switzerland, Germany and the Czech Republic demonstrated that small rodents including *Myodes glareolus*, *Microtus arvalis*, *Microtus agrestis*, *Apodemus flavicollis* and *Apodemus sylvaticus* harbored *A. phagocytophilum* and were suggested as potential reservoirs [24,42-45]. In a study in Northern England, Bown et al. [42] described the maintenance of the enzootic cycle of *A. phagocytophilum* in the rodent-*I. trianguliceps* system. In a study conducted in Germany [43] *A. phagocytophilum* was detected in 13.4% of red bank voles and 6.2% of field voles. In contrast, only 0.5% of *A. flavicollis* was *A. phagocytophilum* positive. Investigations from Switzerland, England and Norway have shown that deer and sheep can be reservoir hosts [14,40,46]. Migrating birds have also been considered important in the dispersal of *A. phagocytophilum* infected *I. ricinus* in Europe and in the distribution of HGA [17,38].

A. phagocytophilum could not be detected in 49 rodents and in 24 *I. ricinus* nymphs feeding on rodents investigated on Jomfruland, possibly because *I. trianguliceps* is the main vector for *Anaplasma* in rodents [24,42]. *A. phagocytophilum* was found in ticks feeding on birds on Jomfruland [47]. This indicates that birds are involved in the maintenance of *Anaplasma* here, but rodents play only a minor role in the epidemiology of *Anaplasma* in the investigated areas in Norway. Hinnebu is located inland and is not on the main route of migrating birds. Tjore is near the coast, but not a typical site for migrating birds. Migrating birds, however, may play an important role as hosts for *I. ricinus* larvae and nymphs and probably for the infection route of *Anaplasma* (as for *B. burgdorferi* s.l.) [47]. On the island of Jomfruland the figures for *A. phagocytophilum* were 8.7%, 11.5% and 23% in 2006-2008. However, *A. phagocytophilum* was found on ticks feeding on birds in 33 out of 308 ticks investigated [47] on Jomfruland and also in questing ticks (Table 4). This indicates that birds are a possible reservoir. Both *B. burgdorferi* s.l. and *A. phagocytophilum* were found in ticks feeding on migrating birds and in questing ticks.

Conclusion

A high prevalence of *A. phagocytophilum* in questing ticks in sites with high abundance of deer (>3 killed animals per km²) and low prevalence of *B. burgdorferi* s.l. was found, and we conclude that deer may be important reservoirs of *A. phagocytophilum* and incompetent carriers for

B. burgdorferi s.l., thereby reducing the infection rate on questing *Ixodes ricinus* ticks.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

OR and AP have designed and performed the experimental study. OR has drafted the manuscript. JR has carried out the statistical and molecular genetic analyses. All authors read and approved the final manuscript

Acknowledgements

We thank the Lithuanian State Science and Studies Foundation and Telemark University College for financial support.

References

- Wang G, van Dam AP, Schwartz I, Dankert J: **Molecular typing of *Borrelia burgdorferi* sensu lato: taxonomic, epidemiological, and clinical implications.** *Clin Microbiol Rev* 1999, **12**:633-653.
- Balmelli T, Piffaretti JC: **Association between clinical manifestations of Lyme disease and different species of *Borrelia burgdorferi* sensu lato.** *Res Microbiol* 1995, **146**:329-340.
- Lane RS, Piesman J, Burgdorfer W: **Lyme borreliosis: Relation of its causative agent to its vectors and hosts in North America and Europe.** *Annu Rev Entomol* 1991, **36**:587-609.
- Wilson ML, Telford SR, Piesman J, Spielman A: **Reduced abundance of immature *Ixodes dammini* (Acari:Ixodidae) following elimination of deer.** *J Med Entomol* 1988, **25**:224-228.
- Kurtenbach K, Sewell HS, Ogden NH, Randolph SE, Nuttall PA: **Serum complement sensitivity as a Key factor in Lyme disease ecology.** *Infect Immun* 1998, **66**:1248-1251.
- Thomas G, Jaenson T, Tällerklint L: **Incompetence of roe deer as reservoirs of the Lyme borreliosis spirochete.** *J Med Entomol* 1992, **29**:813-817.
- Kurtenbach K, De Michelis S, Etti S, Schäfer SM, Sewell HS, Brade V, Kraczy P: **Host association of *Borrelia burgdorferi* sensu lato-the key role of host complement.** *Trends Microbiol* 2002, **10**:74-79.
- Matuschka F-R, Heiler M, Eiffert H, Fisher P, Lotter H, Spielman A: **Diversory role of hoofed game in the transmission of Lyme disease spirochetes.** *Am J Trop Med Hyg* 1993, **48**:693-699.
- Telford SR, Mather TN, Moore SI, Wilson ML, Spielman A: **Incompetence of deer as reservoirs of the Lyme disease spirochete.** *Am J Trop Med Hyg* 1988, **39**:105-109.
- Kurtenbach K, Haninková K, Tsao JI, Margos G, Fish D, Ogden NH: **Fundamental process in the evolutionary ecology of Lyme borreliosis.** *Nat Rev Microbiol* 2006, **4**:660-669.
- Stuen S: **Sjodogg (tick-borne fever) - a historical review.** *Nor Vet Tidsskr* 1998, **110**:703-706. in Norwegian
- Jenkins A, Handeland S, Stuen S, Schouls L, Pol I Van De, Meen RT, Kristiansen BE: **Ehrlichiosis in a moose calf.** *J Wildl Dis* 2001, **37**:201-203.
- Stuen S, Åkerstedt J, Bergström K, Handeland K: **Antibodies to granulocytic Ehrlichia in moose, red deer, and roe deer in Norway.** *J Wildl Dis* 2002, **38**:1-6.
- Liz JS, Sumner JW, Pfister K, Brossard M: **PCR detection and serological evidence of granulocytic ehrlichial infection in roe deer (*Capreolus capreolus*) and chamois (*Rupicapra rupicapra*).** *J Clin Microbiol* 2002, **40**:892-897.
- Chen S-M, Dumler JS, Bakken JS, Walker D: **Identification of granulocytotropic Ehrlichia species as the etiologic agent of human disease.** *J Clin Microbiol* 1994, **32**:589-595.
- Parola P, Davoust B, Raoult D: **Tick- and flea-borne rickettsial emerging zoonoses.** *Vet Res* 2005, **36**:469-492.
- Bjöersdorff A, Berglund J, Kristiansen BE, Söderström C, Eliasson I: **Varying clinical picture and course of human granulocytic ehrlichiosis. Twelve Scandinavian cases of the new tick borne zoonosis are presented.** *Läkartidningen* 1999, **96**:4200-42004. in Swedish
- Mehl R: **The distribution and host relations of Norwegian ticks (Acari, Ixodidae).** *Norw J Entomol* 1983, **30**:46-51.
- Jaenson TGT, Tällerklint L, Lundquist L, Olsén B, Chirico J, Mejlom H: **Geographical distribution, host associations and vector roles of ticks (Acari, Ixodidae & Argasidae) in Sweden.** *J Med Entomol* 1994, **31**:240-256.
- Jääskeläinen AE, Tikkakoski T, Uzcátegui NY, Alekseev AN, Vaheeri A, Vapalahti O: **Siberian subtype Tickborne Encephalitis Virus, Finland.** *Emerg Infect Dis* 2006, **12**:1568-1571.
- Petrovec M, Sumner JW, Nicholson WL, Childs JE, Strle F, Barlič J, Lotrič-Furlan S, Avšič Županc T: **Identity of Ehrlichial DNA sequences derived from *Ixodes ricinus* ticks with those obtained from patients with human granulocytic ehrlichiosis in Slovenia.** *J Clin Microbiol* 1999, **37**:209-210.
- Zhang L, Shan A, Mathew B, Yin J, Fu X, Zhang J, Lu J, Xu J, Dumler S: **Rickettsial seroepidemiology among farm workers, Tianjin, People's Republic of China.** *Emerg Infect Dis* 2008, **14**:938-940.
- Walder G, Lkhamsuren E, Shagdar A, Bataa J, Batmunckh T, Orth D, Heinz FX, Danichova A, Khasnatinov MA, Würzner R, Dierich MP: **Serological evidence for tick borne encephalitis, borreliosis, and human granulocytic anaplasmosis in Mongolia.** *Int J Med Microbiol* 2006, **296**:69-75.
- Bown KJ, Begon M, Bennett M, Woldehiwet Z, Ogden NH: **Seasonal dynamics of *Anaplasma phagocytophila* in a rodent-tick (*Ixodes trianguliceps*) system, United Kingdom.** *Emerg Infect Dis* 2003, **9**:63-70.
- Hillyard R: **Ticks of North-West Europe. Synopsis of the British Fauna (New Series). Volume 58.** Edited by: Barnes RSK, Crothers JH. The Natural Historical Museum, London :178.
- Stańczak J, Racewicz M, Kubica-Biernat B, Kruminis-Łozowska W, Dabrowski J, Adamczyk A, Markowska M: **Prevalence of *Borrelia burgdorferi* sensu lato in *Ixodes ricinus* ticks (Acari, Ixodidae) in different Polish woodlands.** *Ann Agr Environ Med* 1999, **6**:127-132.
- Ambrasienė D, Turčinavičiene J, Vaščilo I, Žygiute M: **The prevalence of *Borrelia burgdorferi* in *Ixodes ricinus* ticks detected by PCR in Lithuania.** *Vet Med Zoot* 2004, **28**:45-47.
- Fukanaga M, Yabuki M, Hamase A, Oliver J, Nakao M: **Molecular phylogenetic analysis of Ixodic ticks based on the ribosomal DNA spacer, internal transcribed spacer, sequences.** *J Parasit* 2000, **86**:38-43.
- Radzijeuskaja J, Indriulytė R, Paulauskas A, Ambrasienė D, Turčinavičiėnė J: **Genetic polymorphism study in *Ixodes ricinus* L. ticks populations of Lithuania using RAPD markers.** *Acta Zool Lith* 2005, **15**:341-348.
- Demaerschalck I, Massaoud A, Kesel M, Hoyois B, Lobet Y, Hoet B, Bigaignon G, Bollen A, Godfroid E: **Simultaneous presence of different *Borrelia burgdorferi* genospecies in biological fluids of Lyme disease patients.** *J Clin Microbiol* 1995, **33**:602-608.
- Cortney JW, Kostelnik LM, Zeidner NS, Massung RF: **Multiplex real-time PCR for detection of *Anaplasma phagocytophilum* 2004. and *Borrelia burgdorferi*.** *J Clin Microbiol* 2004, **42**:3164-3168.
- Jaenson TGT, Tällerklint L: **Incompetence of roe deer as reservoirs of the Lyme borreliosis spirochete.** *J Med Entomol* 1992, **29**:813-817.
- Oliver JH, Stallknecht D, Chandler FW, James AM, McGuire BS, Howarth E: **Detection of *Borrelia burgdorferi* in laboratory-related *Ixodes dammini* (Acari:Ixodidae) fed on experimentally inoculated white-tailed deer.** *J Med Entomol* 1992, **29**:980-984.
- Pichon B, Mousson L, Figureau C, Rodhain F, Perez-Eid C: **Density of deer in relation to the prevalence of *Borrelia burgdorferi* s.l. in *Ixodes ricinus* nymphs in Rambouillet forest, France.** *Exp Appl Acarol* 1999, **23**:267-275.
- Haninková K, Schäfer S, Etti S, Sewell H, Taragelova V, Ziak D, Labuda M, Kurtenbach K: **Association of *Borrelia afzelii* with rodents in Europe.** *Parasitology* 2003, **126**:11-20.
- Humair PF, Gern L: **The wild hidden face of Lyme borreliosis in Europe.** *Microbes Infect* 2000, **2**:915-922.
- Stuen S, Handeland K, Frammarsvik T, Bergström K: **Experimental Ehrlichia phagocytophila infection in red deer (*Cervus elaphus*).** *Vet Rec* 2001, **149**:390-392.
- Stuen S: ***Anaplasma phagocytophilum* - the most widespread tick-borne infection in animals in Europe.** *Vet Res Comm* 2007, **31**(suppl):79-84.

39. Michalski M, Rosenfield C, Erickson M, Selle R, Bates K, Essar D, Massung R: **Anaplasma phagocytophilum in central and western Wisconsin: a molecular survey.** *Parasitol Res* 2006, **99**:694-699.
40. Stuen S, Moun T, Bernhoft A, Sirkka V: **A parietic condition in an Anaplasma phagocytophilum infected roe deer.** *J Wildl Dis* 2006, **42**:170-174.
41. Rosef O, Radzijeuskaja J, Paulauskas A, Haslekås C: **The prevalence of Anaplasma phagocytophilum in host-seeking Ixodes ricinus ticks in Norway.** *Clin Microbiol Infect* 2009, **15**(Suppl 1):.
42. Bown KJ, Begon M, Bennett M, Birtles RJ, Burthe S, Lambin X, Telfer S, Woldehiwet Z, Ogden NH: **Sympatric Ixodes trianguliceps and Ixodes ricinus ticks feeding on field voles (Microtus agretis): Potential for increased risk of Anaplasma phagocytophilum in the United Kingdom?** *Vector-Borne Zoonot Dis* 2006, **6**:404-410.
43. Hartelt K, Pluta S, Oehme R, Kimmig P: **Spread of ticks and tick-borne diseases in Germany due to global warming.** *Parasitol Res* 2008, **103**(Suppl 1):.
44. Hulínská D, Langrová K, Pejčoch M, Pavlásek I: **Detection of Anaplasma phagocytophilum in animals by real-time polymerase reaction.** *APMIS* 2004, **112**:239-247.
45. Liz JS, Anderes L, Sumner JW, Massung RF, Gern L, Rutti B, Brossard M: **PCR detection of granulocytic Ehrlichia in Ixodes ricinus ticks and wild small mammals in western Switzerland.** *J Clin Microbiol* 2000, **38**:1002-1007.
46. Ogden NH, Casey ANJ, Woldehiwet Z, French NP: **Transmission of Anaplasma phagocytophilum to Ixodes ricinus ticks from sheep in the acute and post acute phases of infection.** *Anaplasma phagocytophilum* 2003, **71**:2071-2078.
47. Paulauskas A, Radzijeuskaja J, Rosef O: **Anaplasma in ticks feeding on migrating birds and questing ticks in Lithuania and Norway.** *Clin Microbiol Infect* 2009, **15**(Suppl 1):.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

